Modern mass spec based proteomics

(Because nucleic acids are overrated)

Presentation outline

- What is "proteomics" ?
- Historical overview over development of the technology
- Applications of proteomics
- Data processing and analysis
- Future perspectives

What is proteomics?

Dictionary definition:

- Proteomics is the systematic characterization of all the proteins in an organism, their abundance, localization, structure, modifications, function and interactions.
- Most researchers take a narrower view
 - \circ Protein-protein interactions
 - Quantitative proteomics
 - \circ Functional proteomics
- Various technogogies can be applied
 Our focus: LC-MS/MS

Development of the technology

(From the deflection of "canal rays" to MudPIT)

- Protein mass spectrometry
- Protein separation
- Data analysis





Protein mass spectrometry

- Mass spec
 - Wilhelm Wien (Foundation), 1898
 - $\circ~$ Sir Joseph Thomson (Neon isotopes) , 1913
- Beginning of protein mass spec

 Problem of protein ionization
 Koichi Tanaka (SLD), 1988
 - John Fenn (ESI), 1989





Protein separation

- 2D gel based approaches

 low sensitivity (staining)
 extensive sample handling
 difficult to reproduce
 no sympathy for the gel
- Chromatography based approaces
 Washburn *et al.* (MudPIT), 2001
 - on-line
 - semi quantitative
 - more sensitive
 - high throughput





A state of the art setup

- MudPIT (multi-dimensional protein identification technology)
- Originally developed at Yates lab



Methodological background Quadrupole-TOF (MS/MS)



Operates on either MS or MS/MS mode

Data Analysis

- Reducing raw data to manageable levels.
- Analysis
- Algorythms
- How to estimate the quality of data

Reducing raw data to manageable levels

Preprocessing

- \circ Peak detection, peak labeling, baseline correction
- \circ Data reduction
 - noise removal, smoothing
- Normalization
- Deconvolution

Ion charge state recognition (isotope patterns)
 Peak alignment



Analysis

- Database search, Mann and Yates
- High throughput data
- High noise
- Computationally intense
- Variety of software





Algorithms

Examples:

- SEQUEST (Yates 1995)
- Mascot
- ProLuCID
- Specral network analysis (Bandeira 2007)

SEQUEST

Basic concept published by Yates et al. in 1995.

- Reverse pseudospectral library search.
- Protein sequences analysed sequentially through entire database.
- Preliminary scoring equation:

 $S_{\rm p} = \left(\sum i_{\rm m}\right) n_{\rm i} (1+\beta) \left(1+\varrho\right) / \eta_{\tau}$

- Cross correlation by Fourier transforming gives final score.
- Detects modified amino acids by testing alternative masses for all possible modification sites.
- Descriptive model.





Figure 1. Schematic of the approach used by the computer algorithm to match tandem mass spectra of modified peptides to sequences in the protein database.

Mascot

- Incorporates a probability based implementation of Mowse, molecular weight search.
- Mowse assigns a statistical weight to each peptide match.
- Mowse factor matrix M:

$$\mathbf{m}_{i,j} = \frac{\mathbf{f}_{i,j}}{\left|\mathbf{f}_{i,j}\right|_{\max \text{ in column } j}}$$

• Scoring equation:

Score =
$$\frac{50,000}{M_{Prot} \times \prod_{n} m_{i,j}}$$

- The total score is the absolute probability that the observed match is a random event.
- High score = low probability.
- Presented as -Log(P).
- Probability-based model.

http://www.matrixscience.com/help/scoring_help.html

ProLuCID

- Combines descriptive and probability-based models.
- Binomial probability preliminary scoring.
- Introduces a ProLuCID Z score.
- Algorithm description:
 - Candidate peptides selected from databases based on the precursor mass and peptide mass tolerance.
 - Binomial probability computed for each candidate:

$$P(x \ge m) = \sum_{i=m}^{n} P(x=k) \quad \text{where} \quad p(x=k) = \frac{n!}{k!(n-k)!} p^{k} (1-p)^{(n-k)}$$

XCorr computed with modified cross-correlation algorithm.
 ProLuCID Z score computed:

$$ZScore = \frac{X - \mu}{SD}$$
 where $SD = \sqrt{\frac{\sum_{i=1}^{n} (x_i - \mu)^2}{n-1}}$

Ref. Poster by Tao Xu et al.

De novo sequencing



http://www.astbury.leeds.ac.uk/facil/MStut/mstutorial.htm

Spectral Network analysis

- Described by Bandeira et al. in 2007.
- Combination of *de novo* and spectral alignment techniques.
- Spectral pairs:
 - \circ Overlapping peptides.
 - Modified vs. unmodified peptides.
- Spectral paires usually avoided due to higher running times.
- Generates covering sets of peptides 7-9 aa. long.
 - \circ Most often a single hit in database.
 - \circ Easily found using a hash function.
 - \circ No need for a database comparison.
- Spectral networks.



How to estimate quality of data?

• Compare to scrambled or reversed databases.

- A peptide from the database is scrambled or reversed and compared to the spectral data.
- \circ Has the same aa ratios but different sequences.
- Many scrambled or reversed hits means bad data.

Applications off protein mass spec

- Post translational modifications
- Protein interactions
- Disease genes and Biomarkers
- Stem cell characterization
- Alternative to microarrays
 - mRNA changes may not be physiologically relevant
 mRNA may not be present in tissue of interest (blood)

Future perspectives

- Functional proteomics
- Quantitative proteomics
- Systems biology

 Integration with other -omics datasets
- Standardization of protocols and analysis
 - Databases "ProteomeExpress"
 - The minimum information about a proteomics experiment (MIAPE)

Difficulties and bottlenecks

- Digestion (poor Km, few and inefficenient proteases)
- Peptide separation
- Masking by abundant proteins
 - Difficult to mass spec transcription factors and other low abundant proteins
- Not all peptides fly
- Isomer identification difficult
- There is hope
 - \circ Field is young and moves fast
 - \circ MudPIT setups are becoming commercially available
 - High demand (everybody wants so be friends with the mass spec guy)