Review Summary – BCH370 - Exam 2

Spectroscopy

Fluorescence / Phosphorescence

Fluorescence (~ 10^{-4} sec to 10^{-9} sec) / Phosphorescence (> 10^{-3} sec) FRET (Fluor. Res. Energy Transfer) Eff. = $1/[1 + (R/Ro)^6]$ – needs "spectral overlap"

Radioactivity and Counting

Radioactive decay processes (α / β+ / β- / E.C.); Radioactivity rays (γ-rays)
Half life: A = Ao exp(-kt) where k = ln2/half-life
Measurement of Radioactivity: Geiger Counter / Film / PhosphorImagers / LSC
Liquid Scintillation Counting: Excited solvent / 1° "fluor" / 2° "fluor" / PM

Electrophoresis – transport of charged particle in an electric field.

Theory: $F_{tot} = qE - fv = ma = m(dv/dt) = 0; v = (qE/f)$

f = $6\pi\eta R$ for spheres; η = Viscosity ~ 0.01g/(cm-sec)

Ferguson Plots: electrophoretic mobility reflects both charge and size/shape

Methods: slab / tube / seq. gels / (native; denatured) / Disc. Gel / PAGE / PFGE / IEF / CE

SDS-PAGE (subunit MW) / buffer system / stains; IEF gels / 2D-PAGE

Centrifugation

Theory: $F_{tot} = m_{eff}\omega^2 r - fv = m\omega^2 r(1 - v'\rho) - fv = ma = m(dv/dt) = 0;$ (v' is "v bar") Preparative Methods: RCF / Rotors / Density Gradient: Zonal vs. Isopycnic Methods Analytical Methods / Modern Analytical Ultracentrifuge

Optics: Schlieren ($\alpha = aK(dc/dx)$; Interference ($\Delta J = (aK\Delta c)/\lambda$); Abspt. optics ($A \sim c$) Sedimentation Velocity: $s = v/\omega^2 r = (m(1 - v'\rho)/f)$; $\rightarrow plot (\ln r) vs. t \rightarrow slope = s\omega^2$ Sed, Vel. plus Diffusion: $D = (kT/f) = (RT/N^o f)$; $\rightarrow s = MD(1 - v'\rho)/RT$ Sedimentation Equil.: $lnc_r - lnc_{rm} = [M\omega^2(1 - v'\rho)/(2RT)](r^2 - r_m^2) \rightarrow plot ln c vs. r^2$

Light Scattering: "Static" vs. "Dynamic"

Wavelength >> particle size; Rayleigh (Static) Scattering – i/I_o = $N[8\pi^4\alpha^2 / r^2\lambda^4](1 + \cos^2\theta)$ Raleigh Ratio: $R_{\theta} = (i_{\theta} / I_{\theta})(r^2 / (1 + \cos^2\theta)) = [2\pi^2 n_o^2(dn/dC)^2 / \lambda^4 N_o^2] CM$ or $R_{\theta} = KCM$ KC/ $R_{\theta} = 1/(M^*P(\theta)) + 2 A_2C$; Mean Square Radius (**Rg**) 10 nm to 150 nm Polydispersity (*Mw/Mn*); If normalized, LS = RI for monomer but LS = 2*RI for dimer Dynamic Light Scattering –Hydrodynamic (Stokes) Radius (**R**_h) 1.5 to 1000 nm Experimental (Use of LS and RI); LS = K_{LS}CM(dn/dC)²: **RI** = K_{RI}C(dn/dC) or LS/RI = M[(K_{LS}/K_{RI})(dn/dC)] or M = K'(LS)/(RI) Wavelength << particle size; SAXS → shape information from interference / folding, binding

CD

Terms: CD / plane polarized light vs. circularly polarized light, etc. Special type of spectroscopy - meas. the difference in left and right handed absorbance A(l)- A(r). The instrument: measurements in far UV 180-240 nm (proteins); 180-300 nm (nucleic acids). CD spectra can distinguish types of secondary structure (helix, sheet, r.coil / B-DNA, A-DNA) etc. Applications: Folding / Secondary Structure / Denaturation / Thermal Stability / Binding

Mass Spectrometry

Produce ions / uses electric and magnetic fields to measure the mass
 / charge ratio of the charged particles: Parts: ion source; analyzer; detector
 Source: Electron impact (EI) / Chemical Ionization (CI) / Fast atom bombardment (FAB)
 Field desorption (FD) /Electrospray ionization (ESI) /Laser desorption (LD)
 Analyzer: ions separated according to mass. Quadrupole / Magnetic Sector / TOF

Detector which produces a signal from the separated ions.

Linked Systems: GC/MS; LC/MS; MS/MS

Time-of Flight (TOF) Mass Spectrometer: particles have same kinetic energy KE = zVor (Ze)Es where "Ze" is the charge, "E" the electric field, and "s" the length of the source region before particle is allowed to "drift" to the detector; $(m/Z) - 2eFs(t/D)^2$ or $m = [2eEs(1/D)^2]Zt^2$ (m

$$m/Z$$
 = 2eEs(t/D)², or m = [2eEs(1/D)²] Z

mass = (constant)
$$x Z x t^2$$

Source of "ions" - Applications with Biomacromolecules -

a) Matrix-Assisted Laser Desorption-Ionization (MALDI) / TOF

b) Electrospray Ionization (ESI): nondestructive / microdroplets

c) Capillary Electrophoresis (CE) and ESI: very small samples – femtomole (10⁻¹⁵)

Sequence Analysis Using Mass Spectrometry: MS/MS