

Review of Scale of Biological World

The cell at 1,000,000 X

Review of Protein Structure / Folding

Goals for this review unit:

Protein Structure

- Definitions of primary, secondary, tertiary and quaternary structures
- Common secondary structures
- Phi, Psi (ϕ / ψ) angles / How to read a Ramachandran Plot
- Common terms used to describe protein structure motifs / domains - some examples

Protein Folding / Unfolding (denaturation)

- Energetics / Intra and Intermolecular forces

Prediction of Protein Structure

Early method – Chou / Fasman

Size scale of the biochemical world.

How big is an atom? Length of chemical bond?

Size of a typical bacterial cell? Eukaryotic cell?

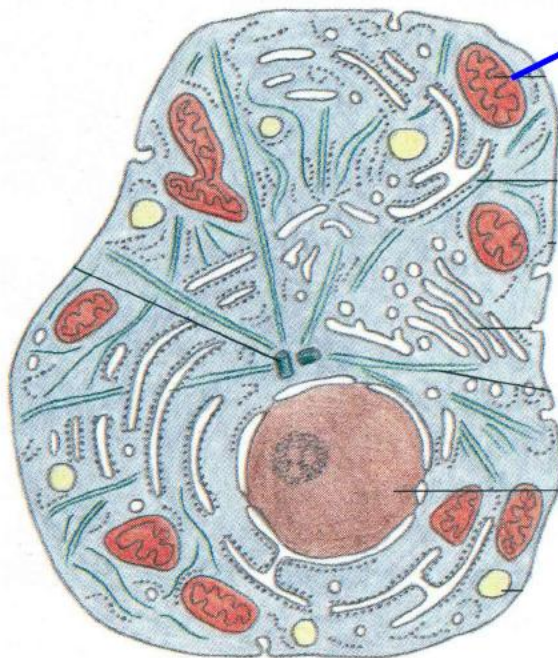


Prokaryotic cells

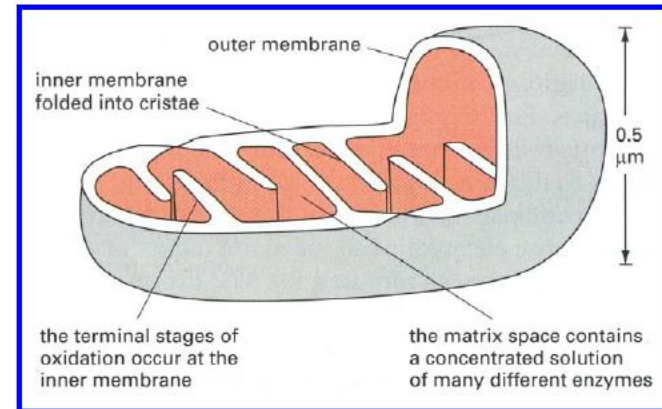


Eukaryotic cells have cell nuclei, other intracellular compartments, many other biological differences from prokaryotes.

Cell components



Mitochondria



- power plants of all eucaryotic cells
- converting food and O_2 into ATP

iClicker: Review Questions

1. What is the approximate diameter of an alpha helix?
A) 0.01 nm B) 0.1 nm C) 1 nm D) 2 nm E) 10 nm
2. Which amino acid has a “phenol” group in its side chain group
A) Trp B) Phe C) Arg D) Lys E) Tyr
3. The K_{eq} for the reaction $A \rightarrow B$ is 10. Under conditions such that the concentration of B is ten times that of A, ΔG° would be expected to be:
A) negative B) zero C) positive

Proteins: Biological function depends on conformation

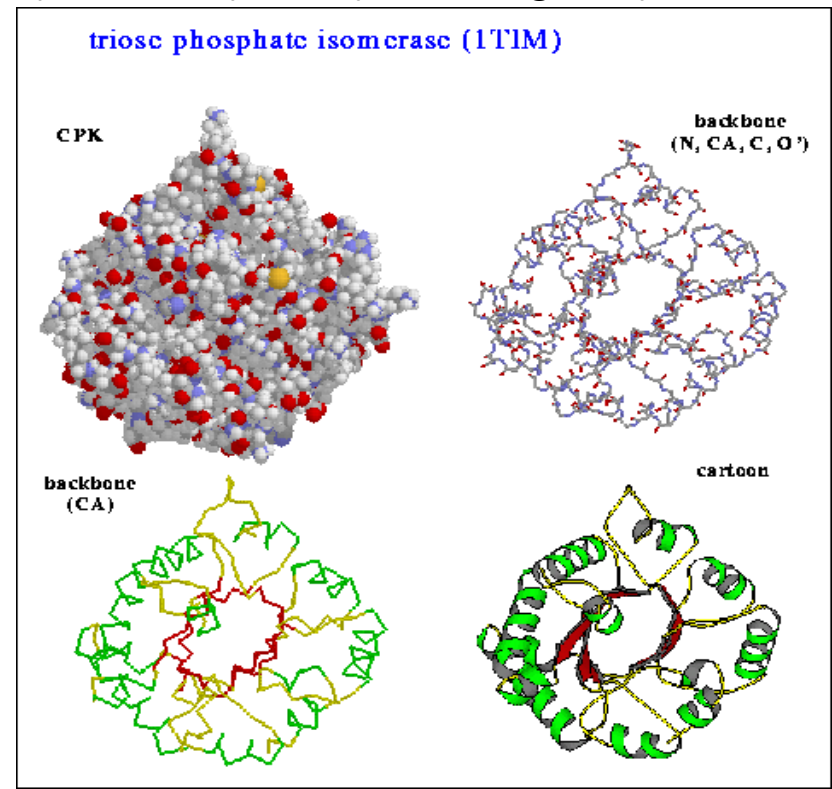
Unique Primary Structure = Unique 3D Structure ??
(Covalent bonds) (Noncovalent Interactions)

Globular Proteins: water soluble, compact, hydrophobic interior / hydrophilic surface
enzymes, receptors, carriers, hormones, etc. (*dynamic agents*)

Fibrous Proteins: water insoluble, structural roles, extended structure
collagen (tendons, bone), α -keratin (hair, nails), etc. (*~static agents*)

APRKEFFVGGNWKMNKSLGELIHTL
NGAKLSADTEVVCGAPSIYLDFARQKL
DAKIGVAAQNCYKVPKGAFTGEISPAM
IKDIGAAWVILGHSERRHVFGESEDELI
GQKVAHALAEGLGVIACIGEKLDEREA
GITEKVVFEQTKAIADNVKDWSKVLA
YEPVWAIGTGKTATPQQAQEVHEKLRG
WLKSHVSDAVAQSTRIIYGGSVTGGNC
KELASQHDVDGFLVGGASLKPEFVDII
NAKH

=



Four Levels of Description of (Native) Protein Structure

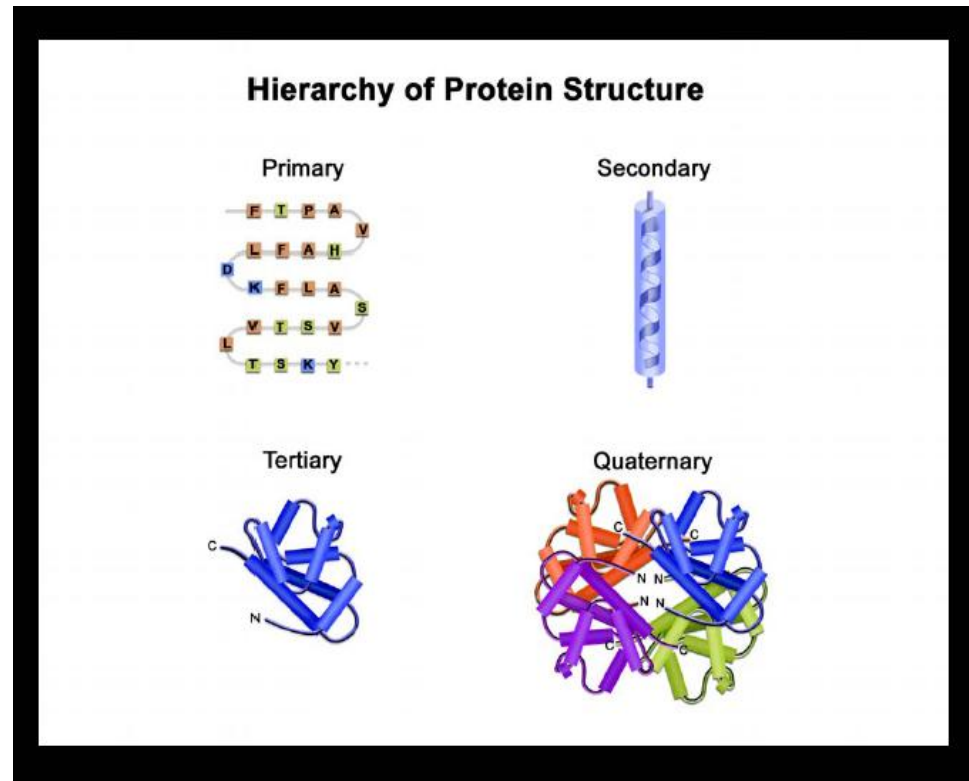
- **Primary Structure:** (~60-1000 amino acid residues) linear seq. of amino acid residues, covalent bonding including -SS- (also called "covalent structure")
*(the **primary structure** of a biological molecule is the exact specification of its atomic composition and the chemical bonds connecting those atoms (including stereochemistry). In general, polypeptides are unbranched polymers. However, proteins can become cross-linked, most commonly by disulfide bonds, and the primary structure also requires specifying the cross-linking atoms, e.g., specifying the cysteines involved in the protein's disulfide or other covalent bonds.)*

- **Secondary Structure:**
Local conformations of backbone, maintained by hydrogen bonds

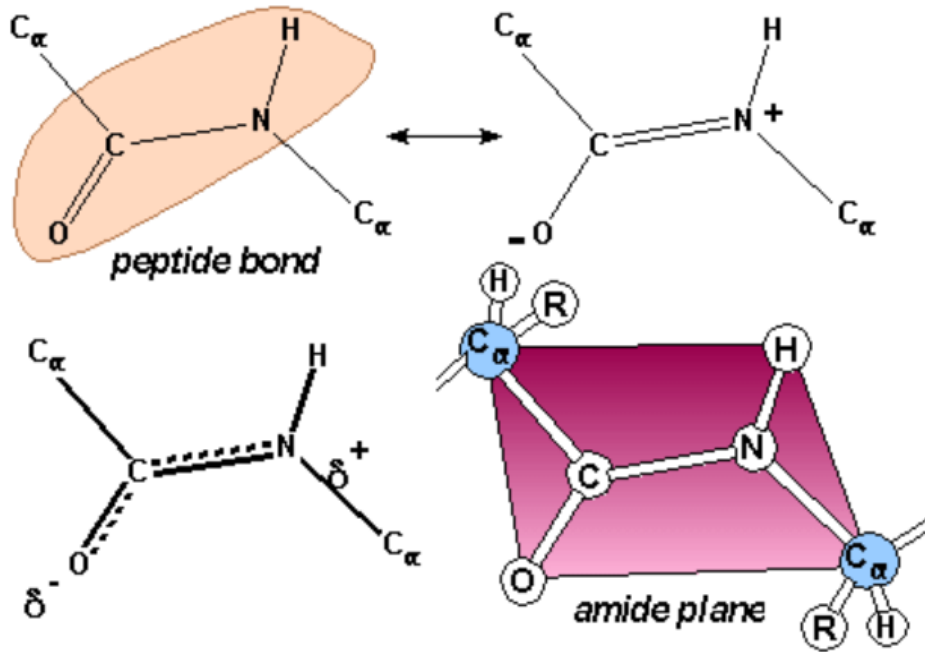
- **Tertiary Structure:**
3D structure of a subunit (one polypeptide chain) in its native state

- **Quaternary Structure:**
Spatial arrangement of subunits in oligomeric proteins

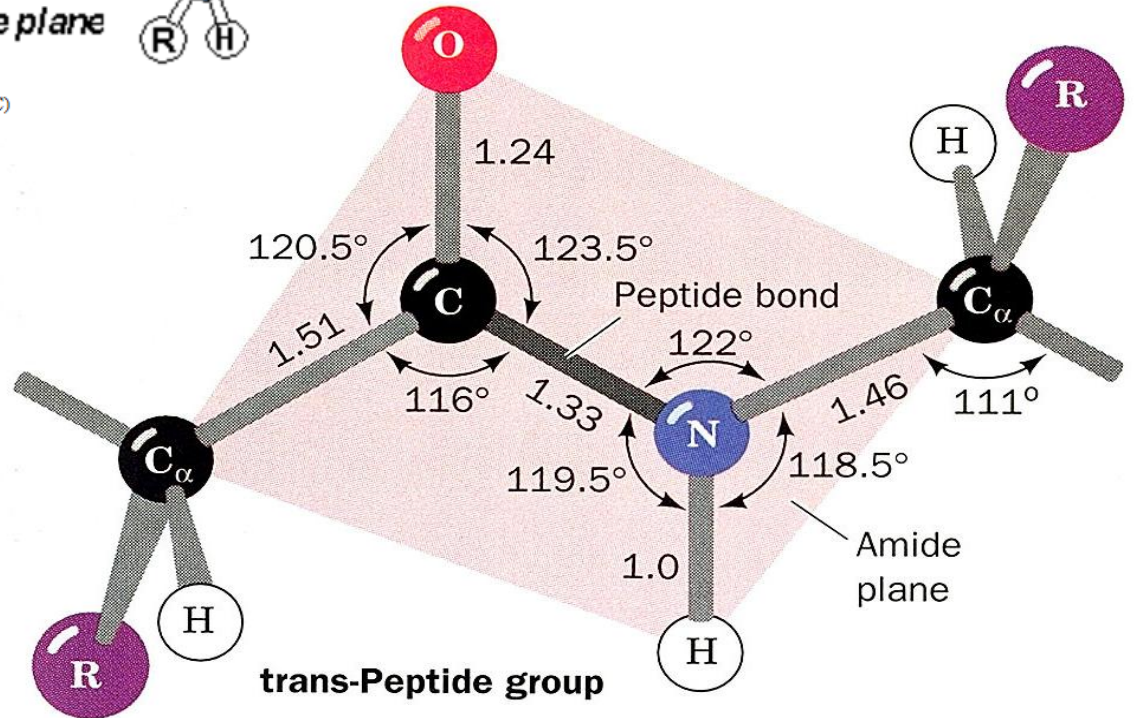
- **Denaturation:** *Partial to complete unfolding*
Denatured Protein: Protein that has lost its native conformation



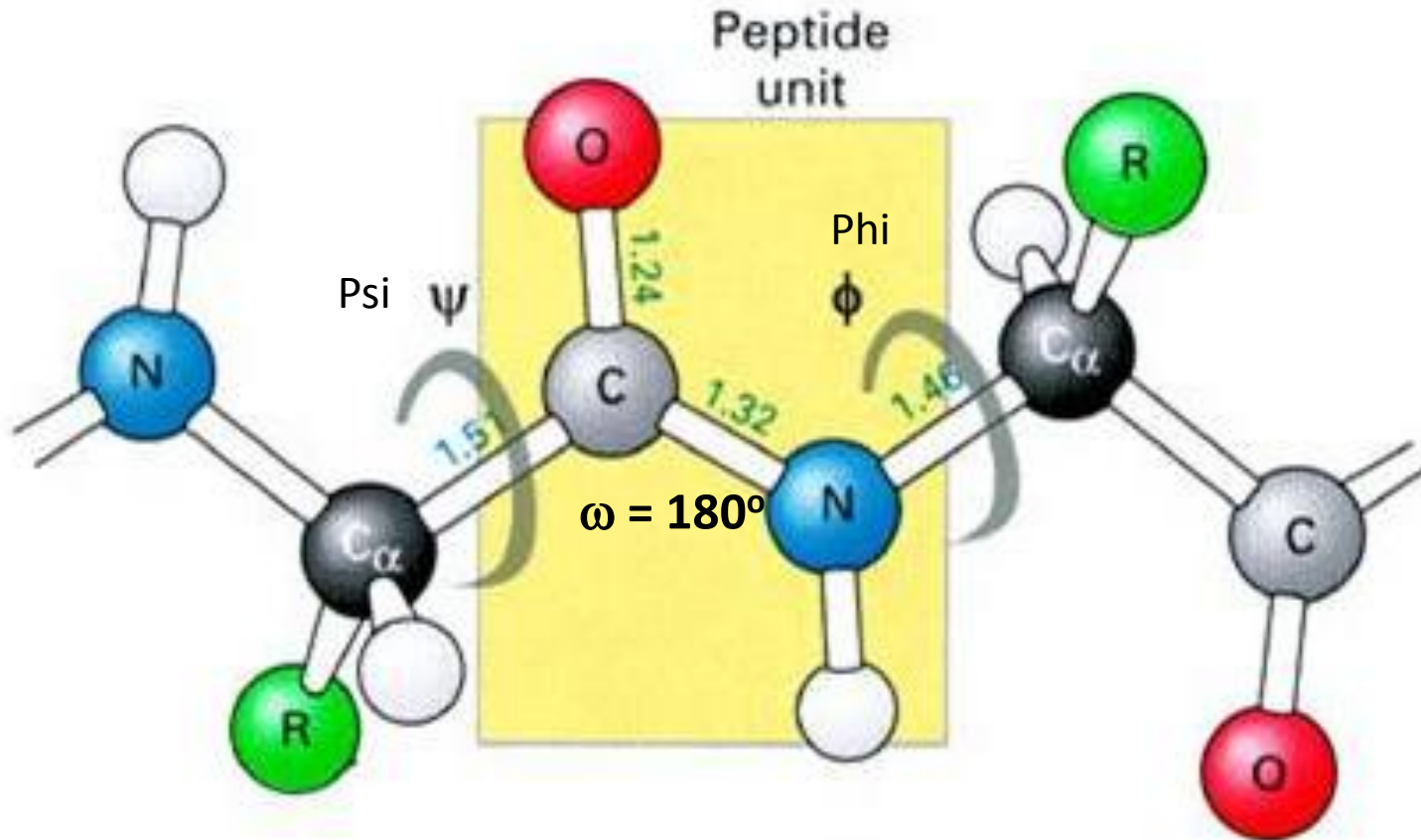
Chemistry of the peptide bond



This image was created by Dr. George Helmkamp, Jr. (UKMC)

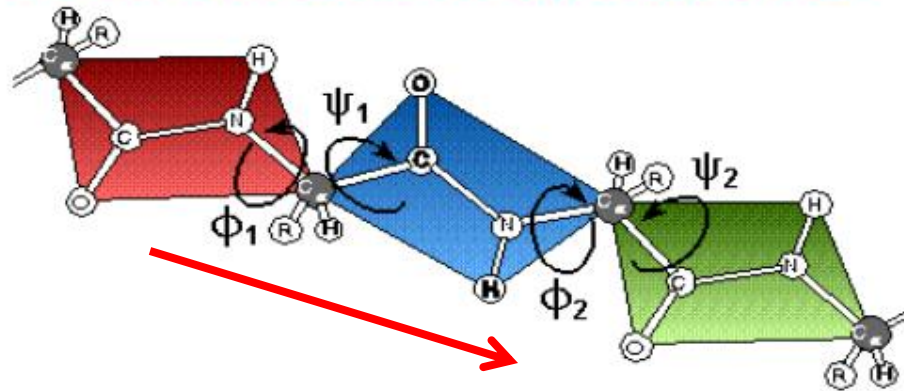


Phi-Psi angles



A peptide has partial double bond character ($\omega = 180^\circ$), thus only two angles (phi and psi) will determine the backbone for trans peptides.

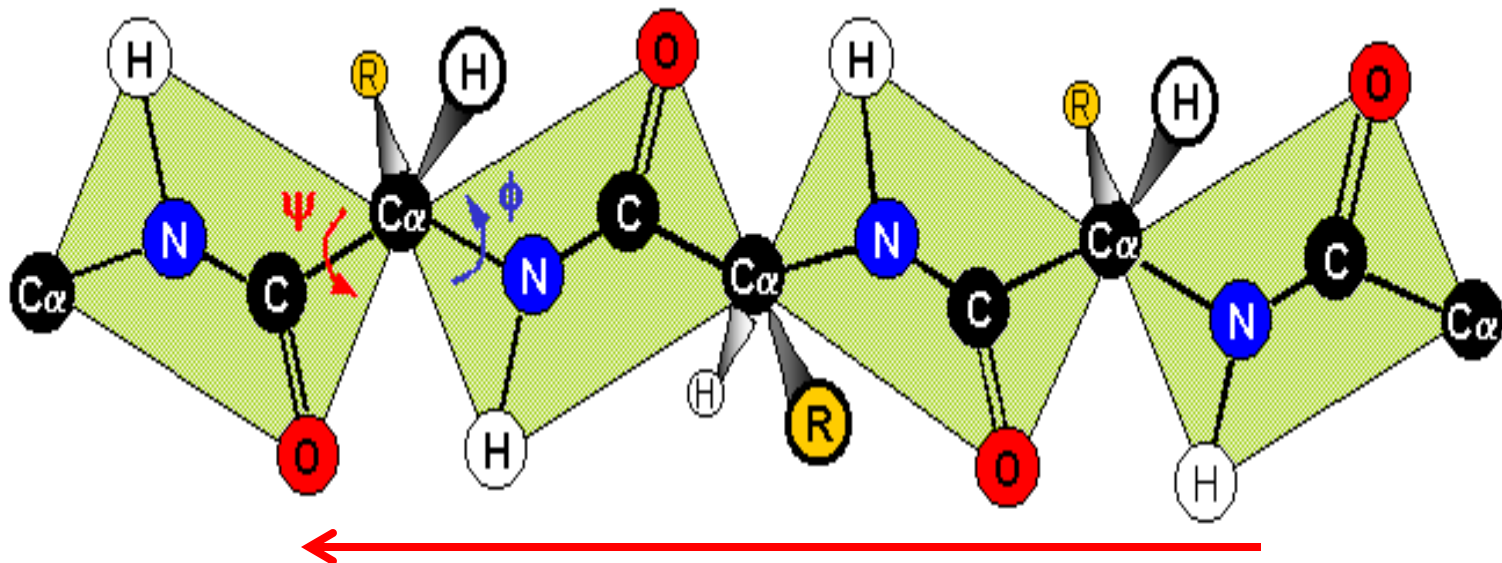
Conformation of a polypeptide



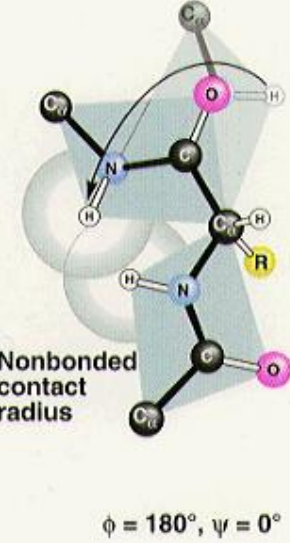
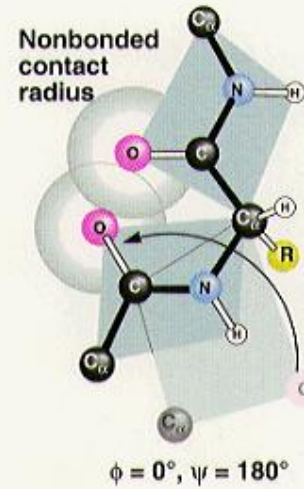
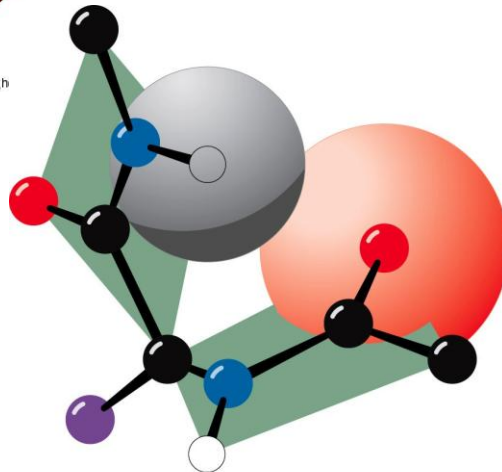
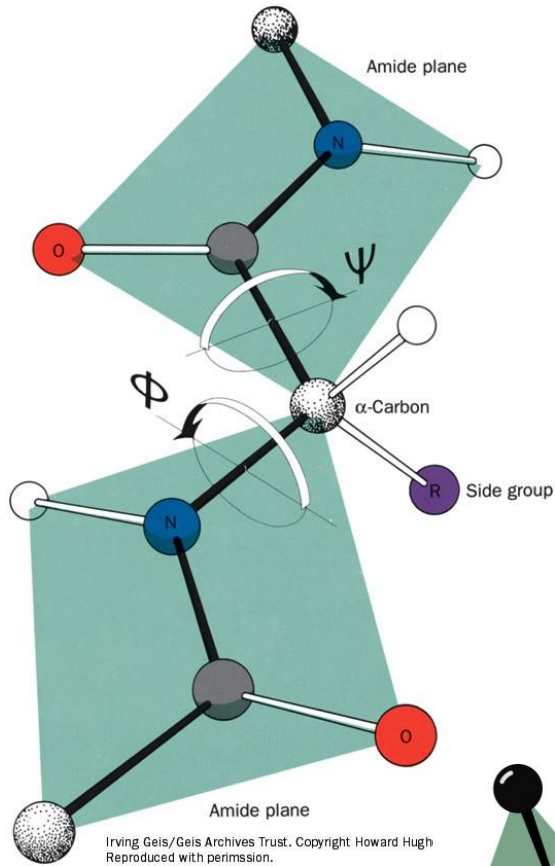
ϕ - rotation around the N-C_α bond

ψ - rotation around the C_α-C bond

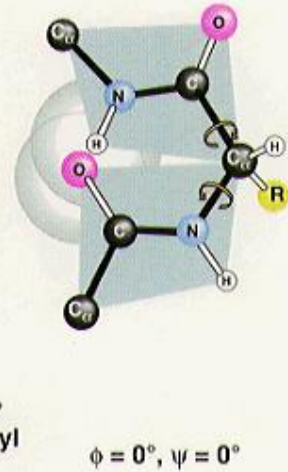
FULLY EXTENDED POLYPEPTIDE CHAIN



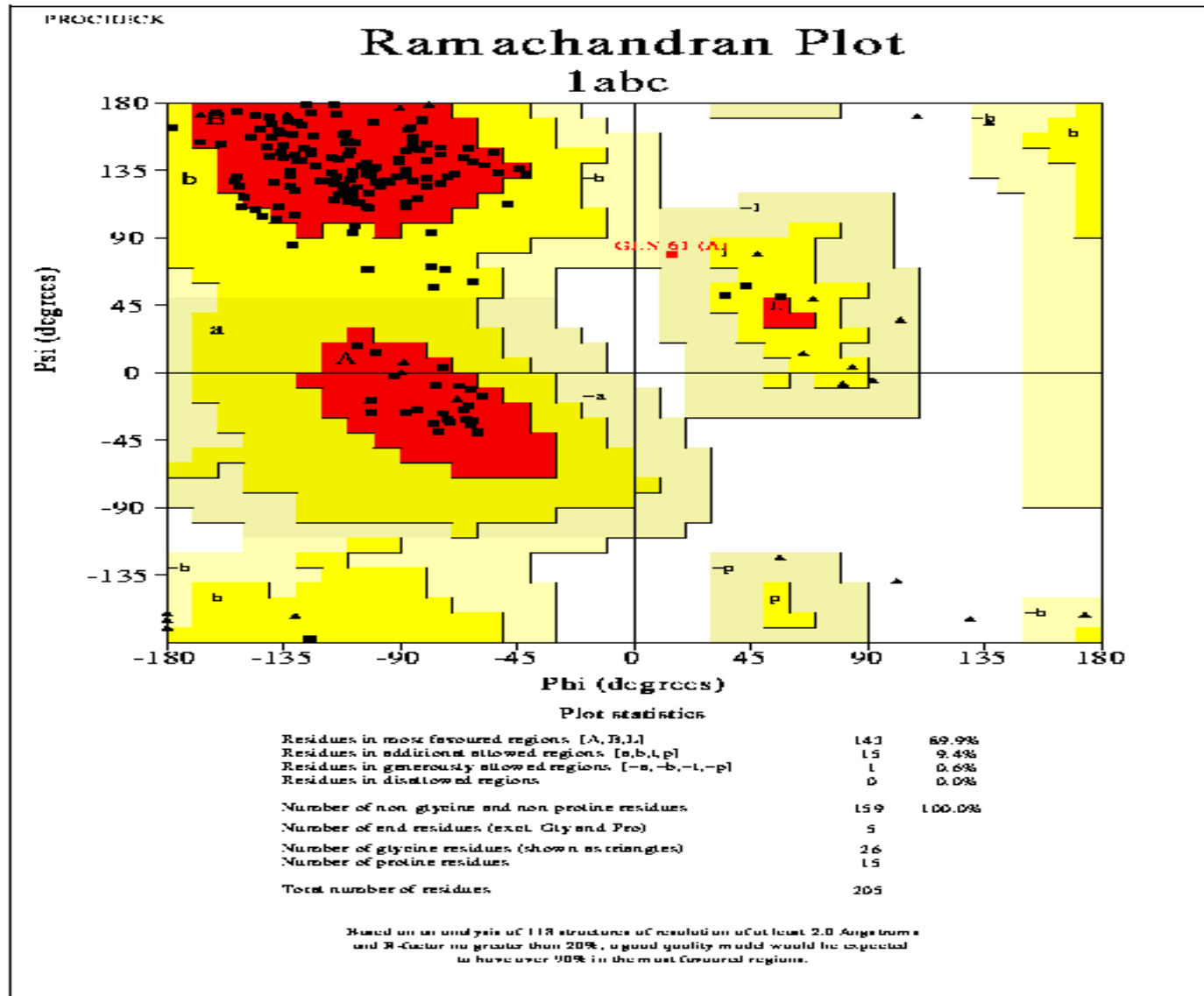
Torsion angles / steric restrictions



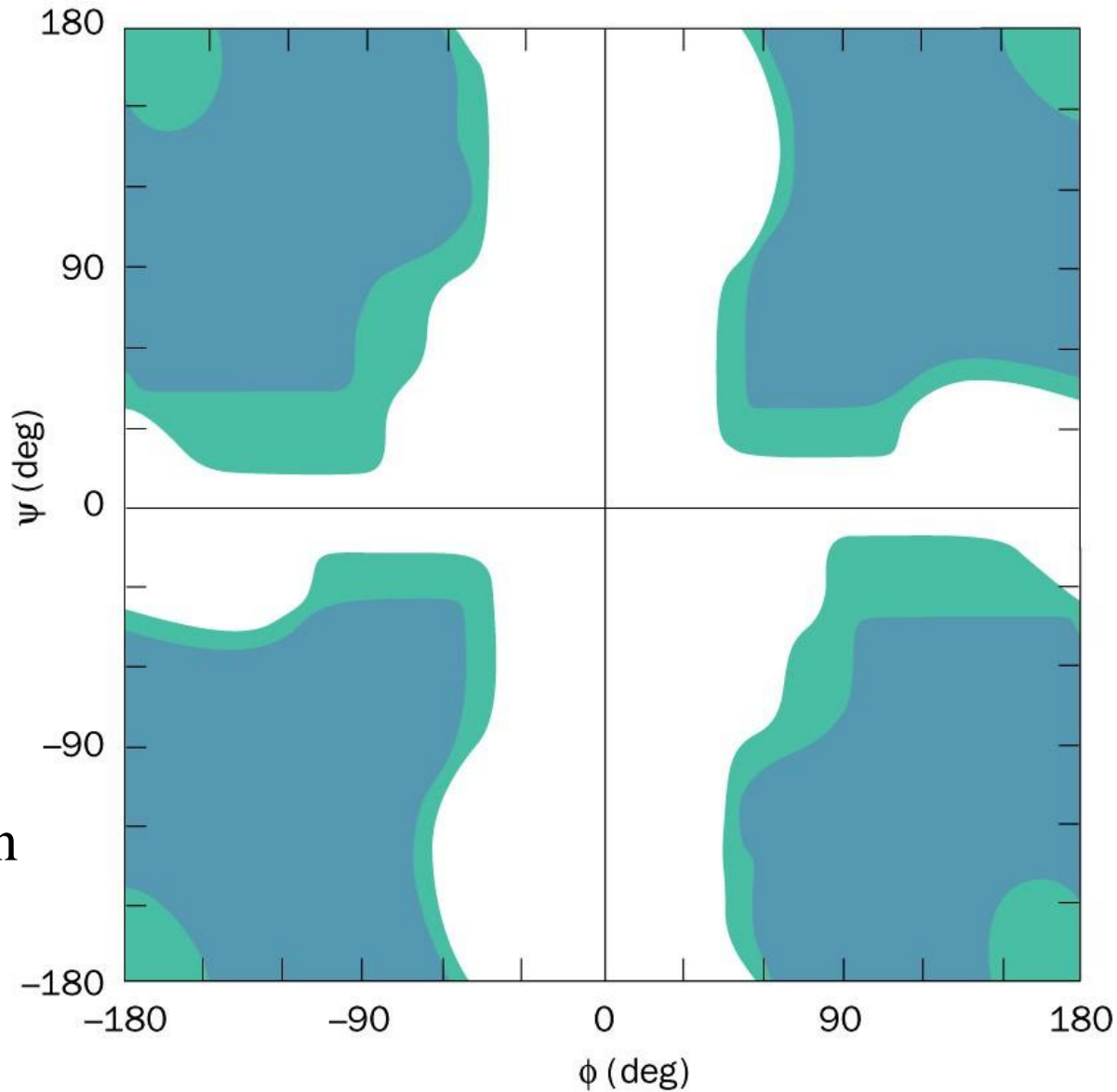
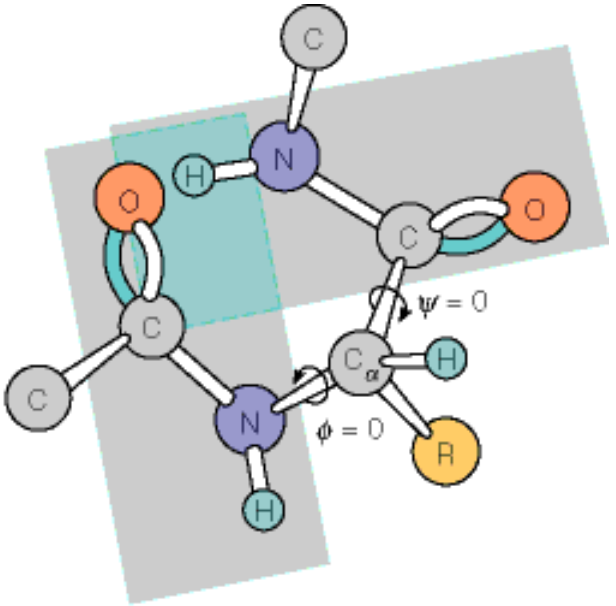
A further ϕ rotation of 120° removes the bulky carbonyl group as far as possible from the side chain



Ramachandran Plot



Allowed torsion angles for Gly residues: Restrictions only from the peptide units



Forbidden conformation in the center of plot

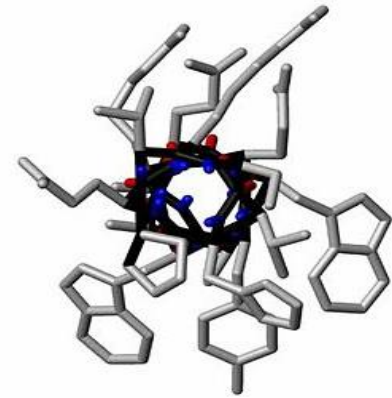
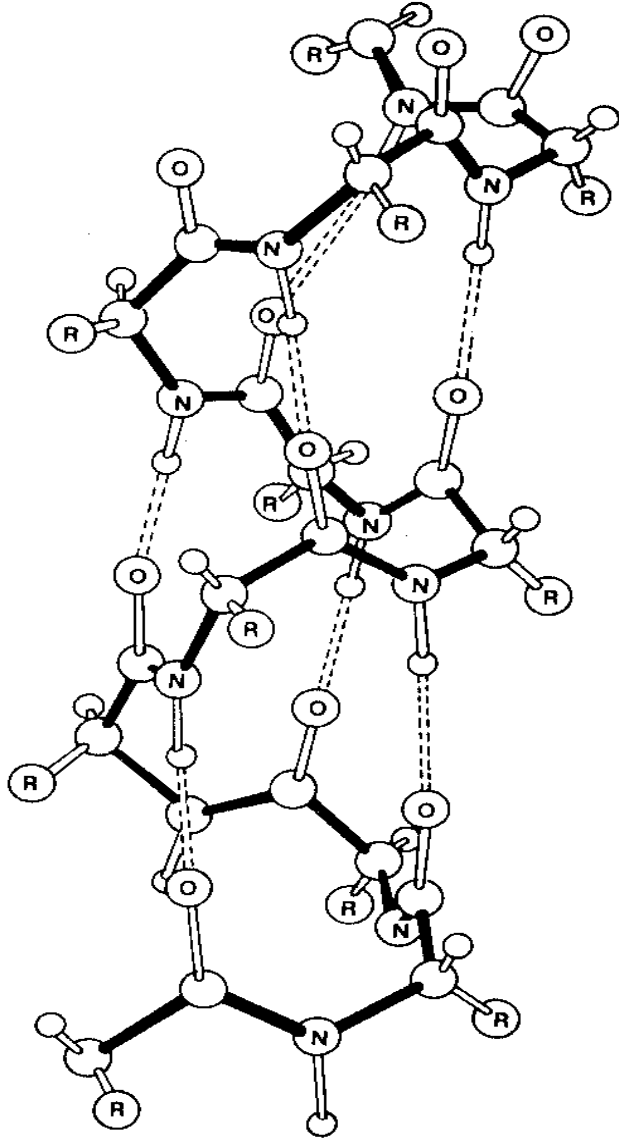
Protein Secondary Structure: Helices

Alpha-helix:

- Right-handed helix
- 3.6 residues per helix turn
- Hydrogen bond between n and $n+4$
- $\phi = -57^\circ$; $\psi = -47^\circ$ (right handed α helix);
- Linus Pauling & Robert Corey - 1951

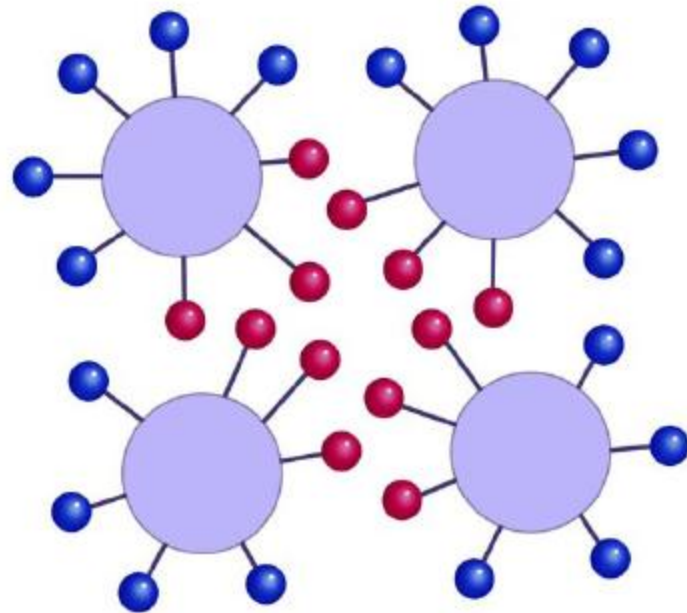
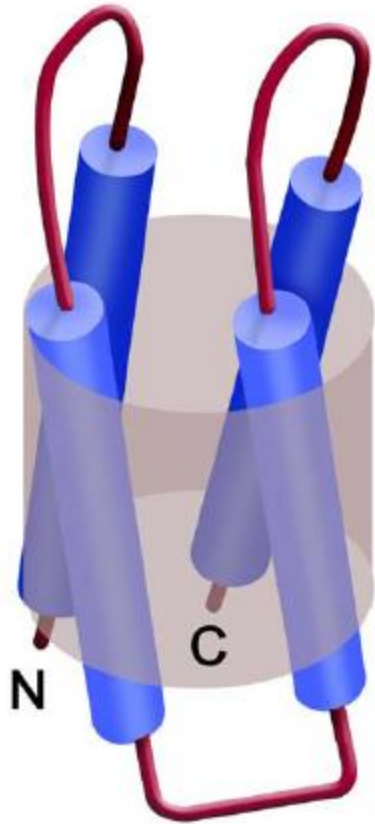
3_{10} helix

- Carbonyl (i) hydrogen bonds to amide (i+3)

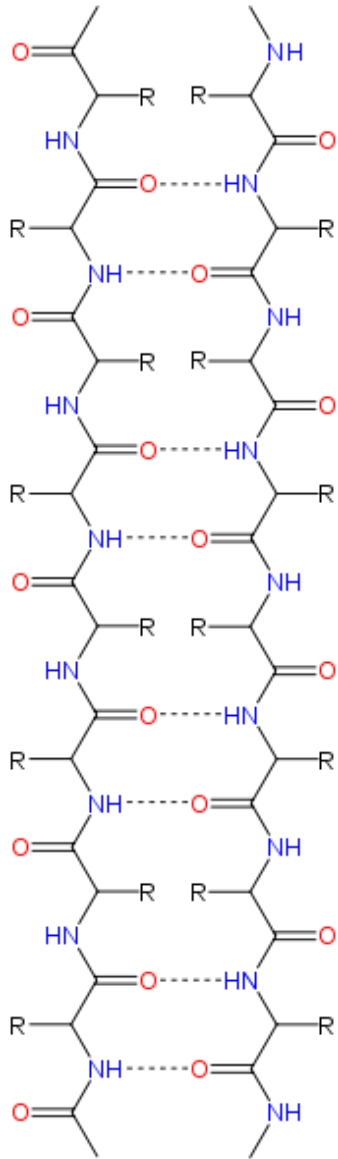


Amphipathic helices

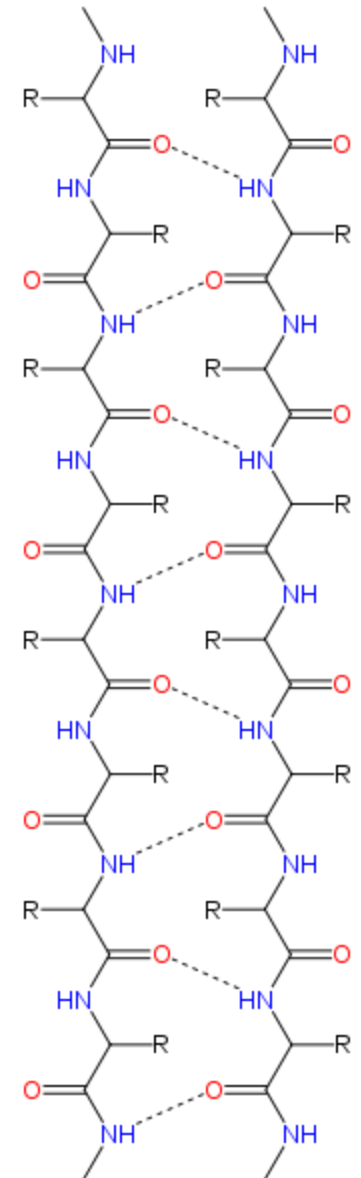
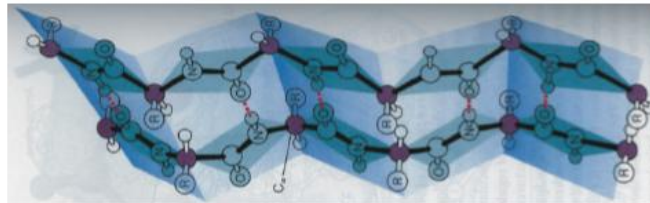
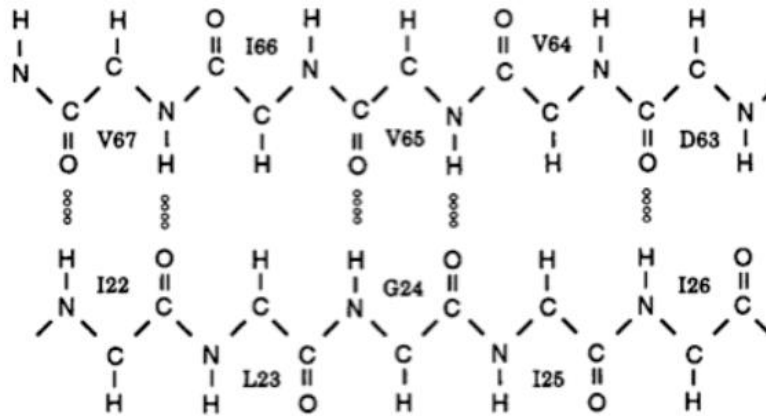
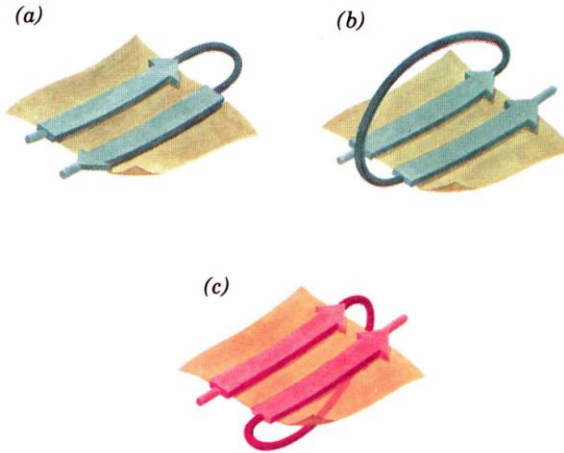
Amphipathic helices



Protein Secondary Structure: Sheet



Anti-parallel



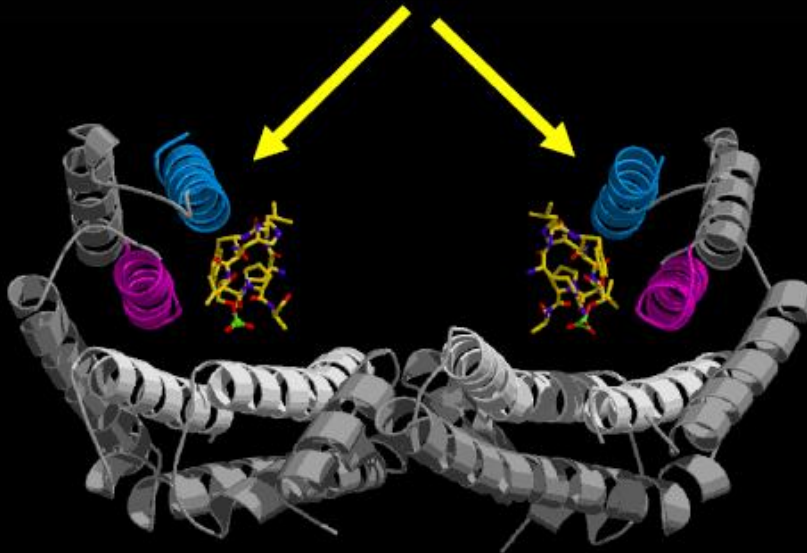
Parallel

Motifs and Domains: Rossmann Fold / Zn finger / Leucine zipper

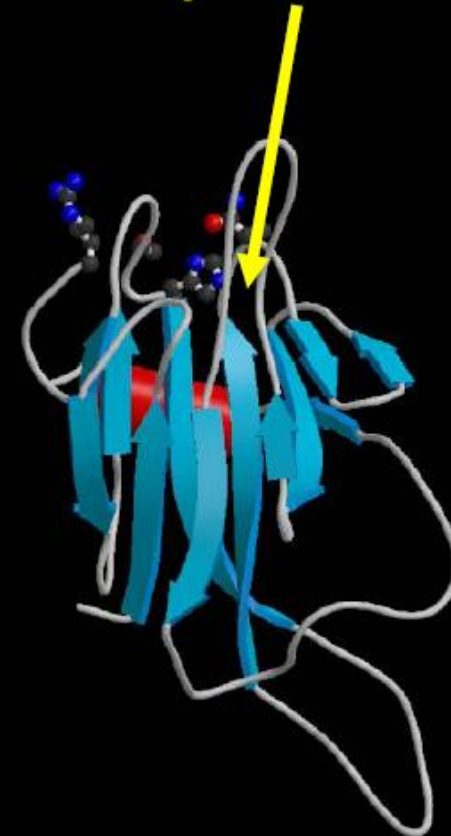
Tertiary Structure: 3D structure

Protein Classes – defined by secondary structural elements

All α -helical



All β -sheet

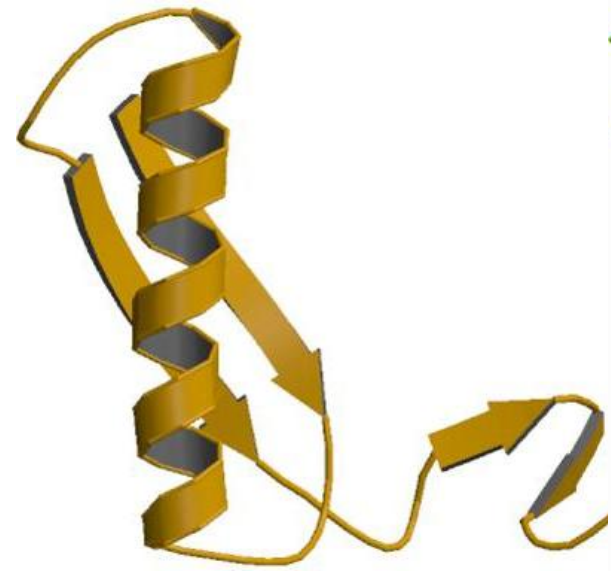


Michael Yaffe

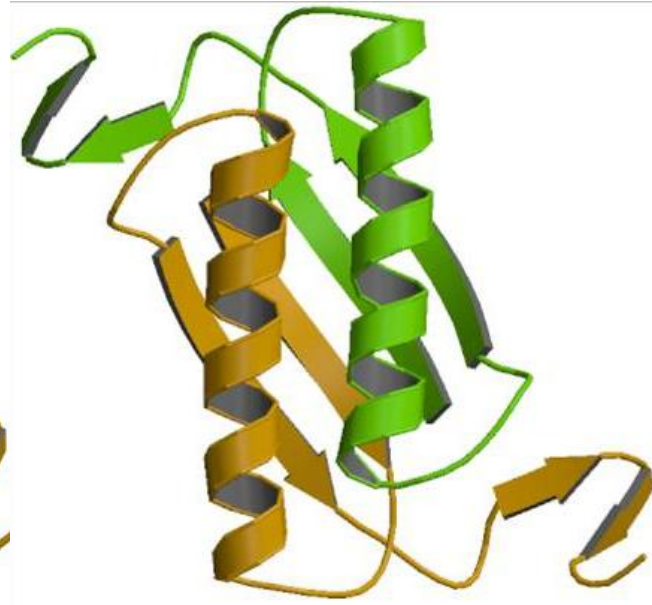
Quaternary Structure:

Arrangements of subunits in oligomers

α_4 ; α_{12} ; $(\alpha\beta)_2$; $(\alpha\beta)_6$



monomer



dimer



Hexamer

4-oxalocrotonate tautomerase

Denaturation (Non-native state): There are many denatured states of macromolecules. Denaturation can occur from many causes:

Denaturation : heat, high salt, hi & lo pH, organic solv., mechanical

- Tm (melting temperature)
- 8M Urea ; 5M guanidinium chloride ; 1% SDS
- Anions : sulfate > phosphate > Cl⁻ > Br⁻ > SCN⁻
- Cations: ammonium > Cs⁺ > K⁺ > Na⁺ > Li⁺ > Mg²⁺ > Ca²⁺ > Ba²⁺

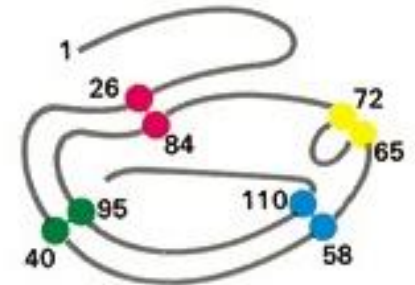
Renaturation :

Chris Anfinsen - Folding of Ribonuclease

124 a.a. + 4 disulfides

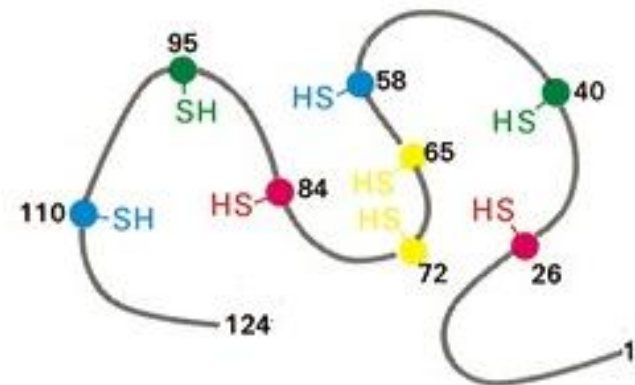
(26 → 84; 40 → 95; 58 → 110; 65 → 72)

(7 x 5 x 3 x 1 = 105 four disulfide combinations)



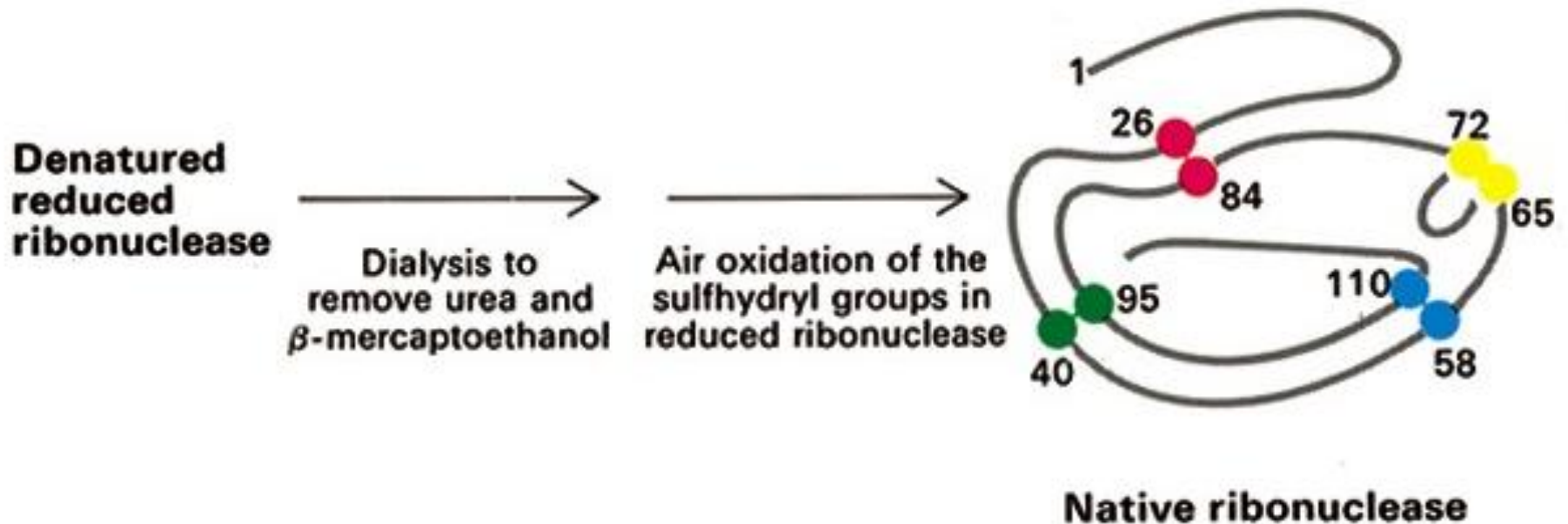
Native ribonuclease

8 M urea and β -mercaptoethanol



Denatured reduced ribonuclease

Chris Anfinsen - Folding of Ribonuclease (4 disulfides)

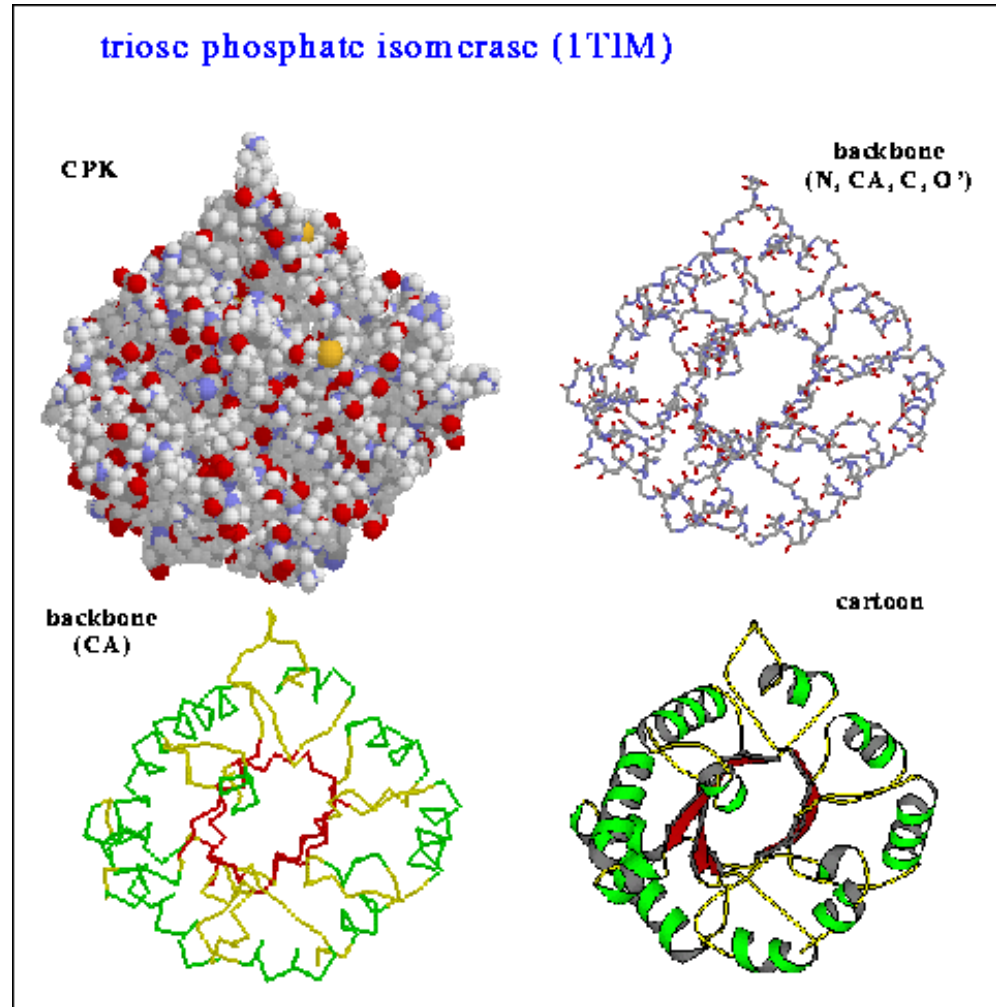


Conclusion: All the information necessary for folding the peptide chain into its native structure is contained in the primary amino acid sequence of the peptide.

Protein Structure

APRKFFVGGNWKMNKDKKSLG
ELIHTLNGAKLSADTEVVCGA
PSIYLDFARQKLDKIGVAAQ
NCYKVPKGAFTGEISPAMIKD
IGAAWVILGHSERRHVFGESE
ELIGQKVAHALAEGLGVIACI
GEKLDEREAGITEKVVFEQTK
AIADNVKDWSKVVLAYEPVWA
IGTGKTATPQQAQEVHEKLRG
WLKSHVSDAVAQSTRIIYGGG
VTGGNCKELASQHDVDGFLVG
GASLKPEFVDIINAKH

=



Protein Folding: Stability / Denaturation and Renaturation

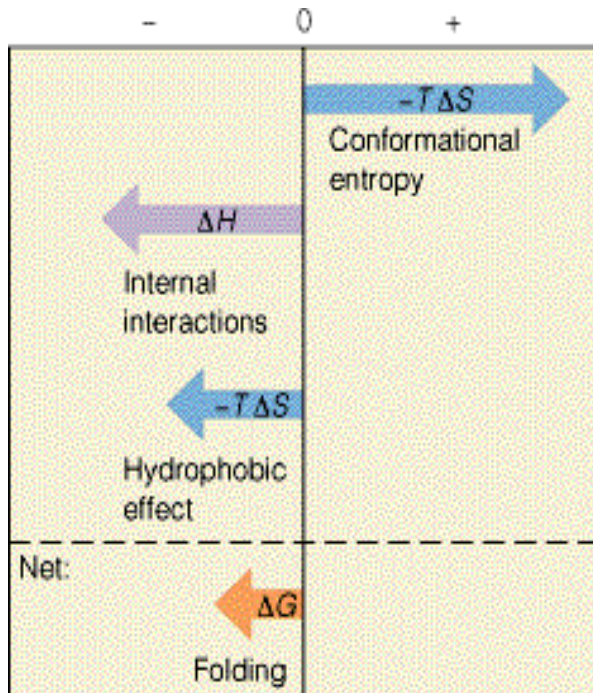
Protein folding (tertiary structure) is determined by weak interactions

H-bonds Dipole interactions Hydrophobic interactions Vander Waals forces Salt bridges

$$\Delta G = G_f - G_u = \Delta H_{\text{prot}} + \Delta H_{\text{solv}} - T\Delta S_{\text{prot}} - T\Delta S_{\text{solv}} \quad (\text{largest } -T\Delta S_{\text{solv}} \text{ for nonpolar R})$$

Folding as a cooperative, sequential process : Local sec. st. / Domains / Molten globules

Molecular chaperones : (GroEL , GroES) assist with folding of some proteins



Protein	ΔG (kJ/mol)	ΔH (kJ/mol)	ΔS (J/K·mol)
Ribonuclease	-46	-280	-790
Chymotrypsin	-55	-270	-720
Lysozyme	-62	-220	-530
Cytochrome c	-44	-52	-27
Myoglobin	-50	0	+170

Note: Data adapted from P. L. Privalov and N. N. Khechinashvili, *J. Mol. Biol.* (1974) 86:665–684. Each data set has been taken at the pH value where the protein is maximally stable; all are near physiological pH. Data are for the folding reaction: Denatured \rightleftharpoons native.

Force that destabilizes protein: Entropy

A folded protein is limited to a much smaller conformation space than an unfolded protein.

Consider backbone only

For an unfolded protein of 100 residues

Each residue: three possible ψ and three possible ϕ

$$S = R \ln W = R \ln 9^{99} = 1.8 \text{kJ/mol} \cdot \text{K}$$

For a folded protein of 100 residues

Each residue: one possible ψ and one possible ϕ

$$S = R \ln W = R \ln 1^{99} = 0$$

$$\Delta G_{\text{conformation}} = -T\Delta S = 540 \text{kJ/mol}$$

Forces that stabilize proteins:

H-bond

Ion-ion interaction

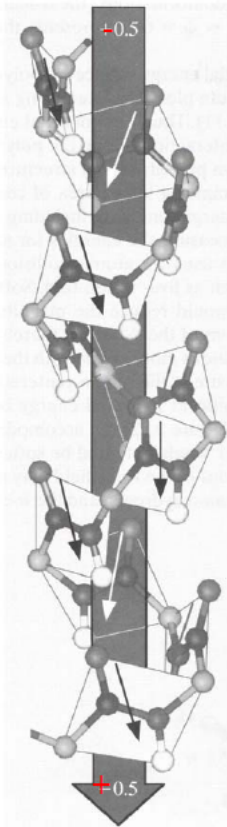
Dipole-related interaction

van de Waals interaction

Hydrophobic interaction

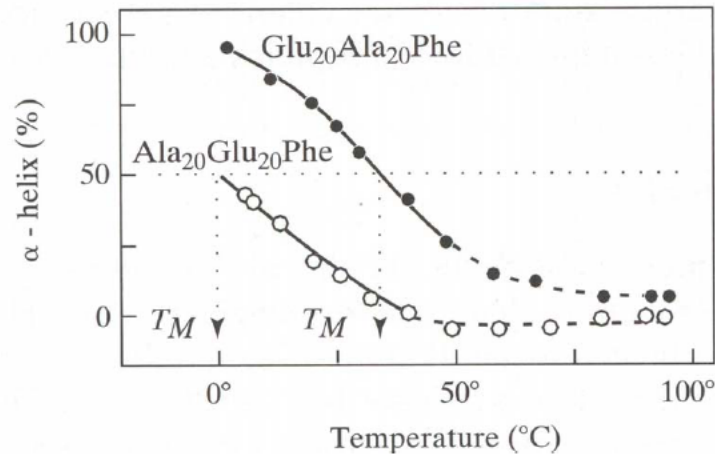
Disulfide-bond

Dipole-related interaction



α -helix has a large dipole moment.

Direction of the dipole: C-terminus to N-terminus



Ala: form a helix

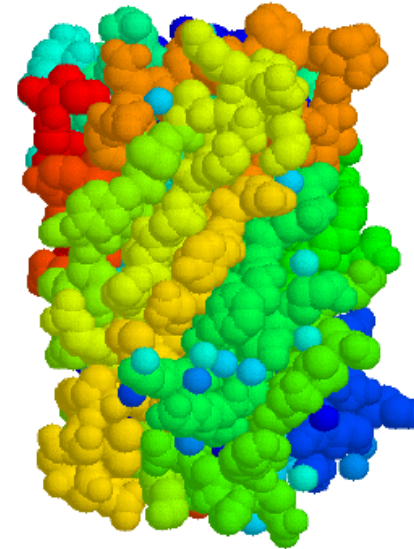
Glu: negatively charge and disordered

Glu₂₀Ala₂₀ is stabilized by the dipole-charge interaction.

Protein folding



sequence



structure

- ◆ Proteins assume specific 3D structures.
- ◆ Protein structures are essential for their function.
- ◆ The protein structures are determined by their sequences.

The Levinthal Paradox (1969)

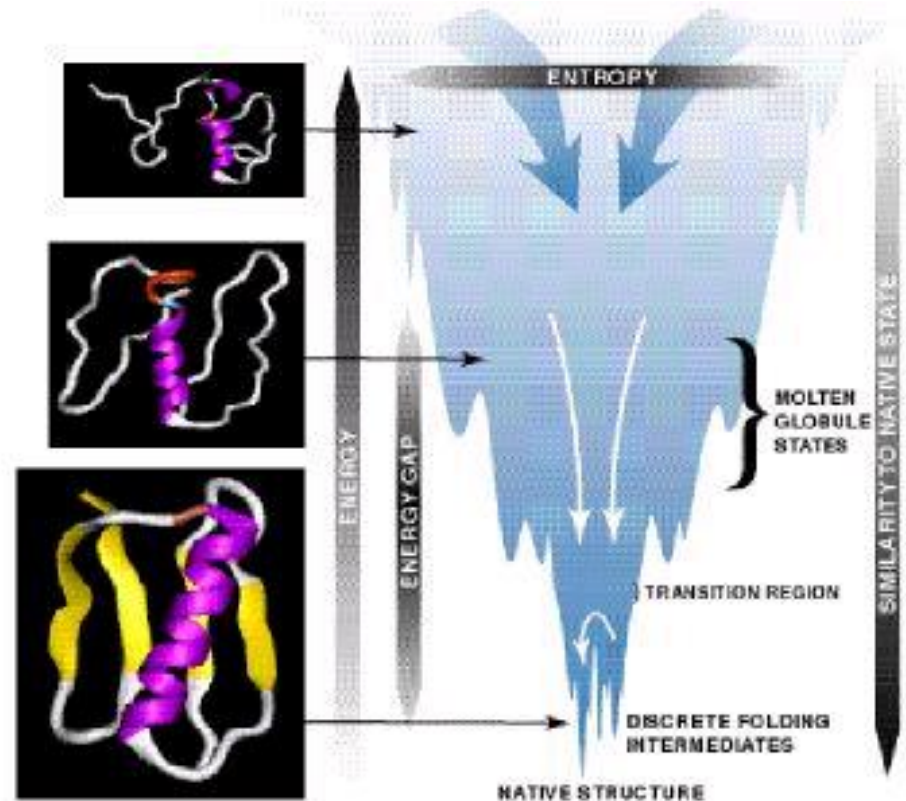
- There are too many possible conformations for a protein to fold by a random search.
- Consider just for the peptide backbone, there are at least 3 conformations per amino acid in the unfolded state, For a 100 a.a. protein we have 3^{100} conformations.
- If the chain can sample 10^{12} conformations/sec, it takes 5×10^{35} sec (2×10^{28} year)
- Conclusion: Protein folding is not random, must have pathways.

Protein Folding Landscape Theory

(Wolynes, Onuchic, Dill, Chan, Sali, Karplus, Brooks etc)

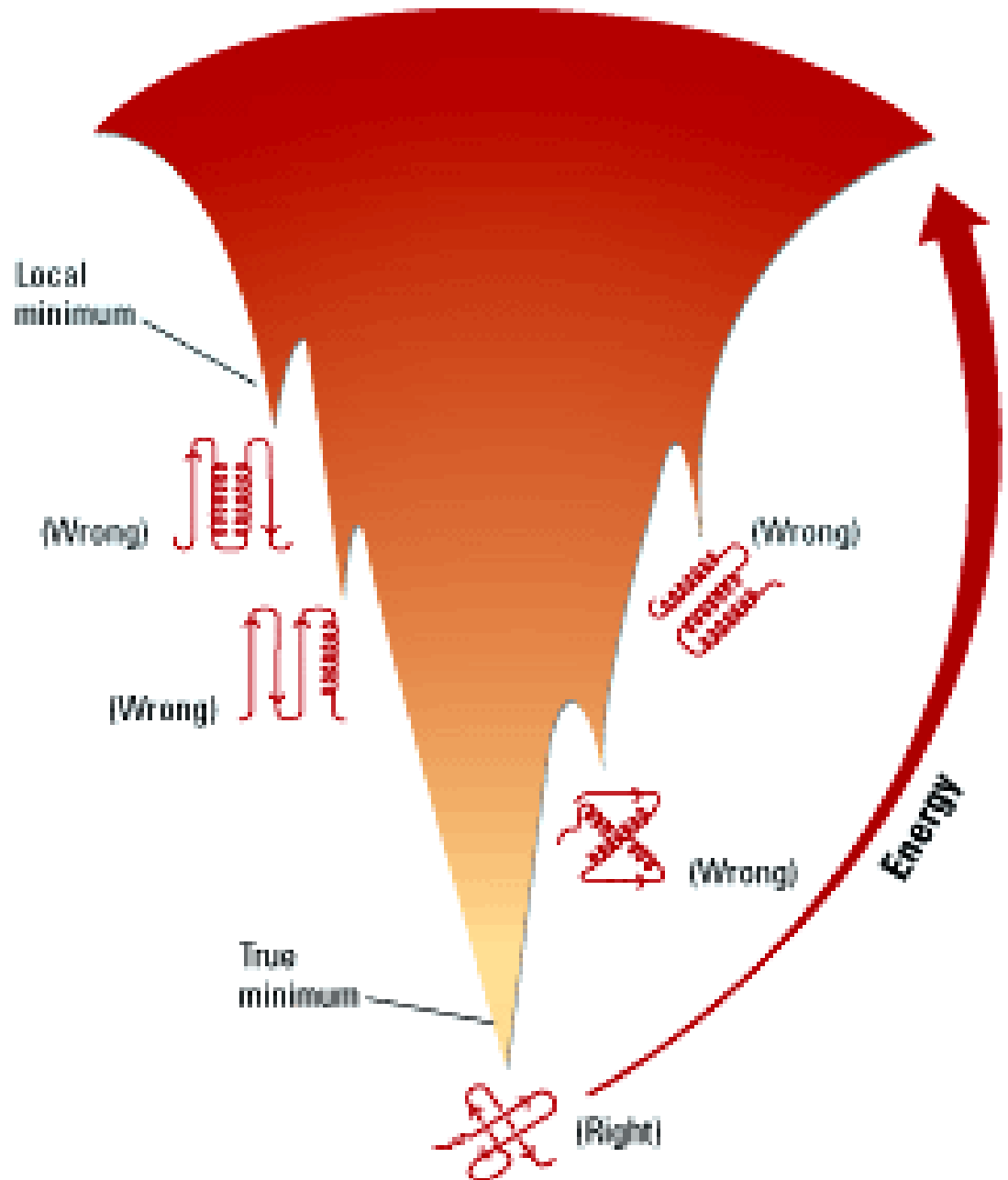
Proteins fold on timescales ranging from a microsecond to a few minutes, so they obviously drive or are driven quickly toward the native state.

- Folding can be described as the descent of the folding chain down a 'folding funnel,' with local roughness of the funnel reflecting the potential for transient trapping in local minima and the overall slope of the funnel representing the thermodynamic drive to the native state.
- A key notion is, in all but the final stages of folding, there exists an ensemble of structures (**molten globules**)--protein folding consequently occurs via **multiple pathways**.



So theoretically, if we have the protein sequence, we can know its structure and its function.

The transition state is composed of a broad ensemble of structures rather than one particular structure.



Chou-Fasman

Biochemistry, 13: 222-245, 1974

- **Statistical Method**

- **Based on 15 proteins of known conformation, 2473 total amino acids**
- **Determined “protein conformational parameters”**
 $P\alpha$, $P\beta$, based on $f_i^s / (\sum f_j^s / 20) \rightarrow 0.5-1.5$

Helical residues		P_α			P_β	β-Sheet residues			
Helical residues	Glu	1.53	H_α	Strong helix former	H_β	Strong sheet former	Met	1.67	
	Ala	1.45					Val	1.65	
	Leu	1.34					Ile	1.60	
	Helical residues	His	1.24	h_α	Helix former	h_β	Sheet former	Cys	1.30
		Met	1.20					Tyr	1.29
		Gln	1.17					Phe	1.28
		Trp	1.14					Gln	1.23
		Val	1.14					Leu	1.22
		Phe	1.12					Thr	1.20
		Trp	1.19					Trp	1.19
Helical residues	Lys	1.07	l_α	Weak helix former	l_β	Weak sheet former	Ala	0.97	
	Ile	1.00					Arg	0.90	
	Helical residues	Asp	0.98	i_α	Helix indifferent	i_α	Sheet indifferent	Gly	0.81
		Thr	0.82					Asp	0.80
		Ser	0.79					Lys	0.74
		Arg	0.79					Ser	0.73
		Cys	0.77					His	0.71
	Helical residues	Asn	0.73	b_α	Helix breaker	b_β	Sheet breaker	Asn	0.65
		Tyr	0.61					Pro	0.62
		Pro	0.59					Glu	0.26
Gly		0.53							
			B_α	Strong helix breaker	B_β	Strong sheet breaker			

Chou-Fasman

Empirical rule set for secondary structure nucleation using $\langle P_\alpha \rangle$, $\langle P_\beta \rangle$

- Search for helical nuclei: locate clusters of 4 (H_α or h_α) out of 6 residues. Unfavorable if $> 1/3$ (b_α or B_α).
- Extend helical segments in both directions until terminated by tetrapeptides with $\langle P_\alpha \rangle < 1.0$. Helix breakers include b_4 , b_{3i} , etc. Some of the tetrapeptide residues can be in the helical ends (except Pro).
- Refine boundaries: Pro, Asp, Glu prefer N-terminal end, His Lys, Arg prefer C-terminal end.
- **Rule #1 – Any segment ≥ 6 residues with $\langle P_\alpha \rangle \geq 1.03$ and $\langle P_\alpha \rangle > \langle P_\beta \rangle$, satisfying above conditions is predicted as helical.**

Chou-Fasman

Empirical rule set for secondary structure nucleation using $\langle P_\alpha \rangle$, $\langle P_\beta \rangle$

- Search for β -sheet nuclei: locate clusters of 3 β residues (H_β or h_β) out of 5 residues. Unfavorable if $> 1/3$ β breakers (b_β or B_β).
- Extend β -sheet segments in both directions until terminated by tetrapeptides with $\langle P_\beta \rangle < 1.0$. β -sheet breakers include b_4 , b_{3i} , etc.
- Refine boundaries: Glu occurs rarely in β -region and Pro equally uncommon within inner β -sheets. Charged residues rare at either end. Trp most frequently at N-terminal end
- **Rule #2 – Any segment ≥ 5 residues with $\langle P_\beta \rangle \geq 1.05$ and $\langle P_\beta \rangle > \langle P_\alpha \rangle$, satisfying above conditions is predicted as β -sheet.**

Predict the secondary structure

Predicted α -Helices $\langle P_\alpha \rangle$	<div style="border: 1px solid black; padding: 2px; display: inline-block;"> h_α H_α H_α I_α H_α I_α 1.20 1.53 1.53 1.70 1.34 1.07 </div> I_α i_α <div style="border: 1px solid black; padding: 2px; display: inline-block;"> I_α I_α I_α h_α h_α h_α 1.07 1.00 1.00 1.12 1.14 1.14 </div> B_α B_α B_α B_α i_α B_α I_α B_α i_α h_α 0.53 0.53 0.59 0.53 0.79 0.53 1.07 0.53 0.82 1.17
	1.29
Sequence	Ac-Met-Glu-Glu-Lys-Leu-Lys-Lys-Ser-Lys-Ile-Ile-Phe-Val-Val-Gly-Gly-Pro-Gly-Ser-Gly-Lys-Gly-Thr-Gln
Observed Structures:	<div style="border: 1px solid black; padding: 2px; display: inline-block; margin-right: 20px;">α-Helix</div> <div style="border: 1px solid black; padding: 2px; display: inline-block; margin-right: 20px;">β-Sheet</div> <div style="border: 1px solid black; padding: 2px; display: inline-block;">Reverse Turns</div>
Predicted β -Sheets $\langle P_\beta \rangle$	H_β B_β B_β b_β h_β b_β b_β b_β b_β <div style="border: 1px solid black; padding: 2px; display: inline-block;"> H_β H_β h_β H_β H_β 1.60 1.60 1.28 1.65 1.65 </div> i_β i_β b_β i_β b_β i_β b_β i_β h_β h_β 1.67 0.26 0.26 0.74 1.22 0.74 0.74 0.72 0.74 0.81 0.81 0.62 0.81 0.72 0.81 0.74 0.81 1.20 1.23
	1.56
Predicted Reverse Turns	1.9 -3.5 -3.5 -3.9 3.8 -3.9 -3.9 0.8 -3.9 4.5 4.5 2.8 4.2 4.2 -0.4 <div style="border: 1px solid black; padding: 2px; display: inline-block;"> -0.4 -1.6 -0.4 -0.8 -0.4 -3.9 -0.4 </div> -0.7 -3.5
	Hydropathy

The reliability of this prediction approach is only 70% because the tertiary structures are not taken into account.

JPred: a consensus secondary structure prediction server

*James A. Cuff^{1,2}, Michele E. Clamp², Asim S. Siddiqui¹,
Matt Finlay¹ and Geoffrey J. Barton^{1,2}*

Uses 6 different prediction methods: DSC, PHD, NNSSP, PREDATOR, MULPRED and ZPRED. Each method is run and the results are combined into a single, consensus structure prediction.

Predictions for request hmg1

```
OrigSeq      : 1-----11-----21-----31-----41-----51-----61-----71-----81-----91-
              : MAAMRKALPRRLVGLASLRVSTSSMGTLPKRVKIVEVGPRLDGLQNEKNIVSTPVKIKLIDMLSEAGLSVIETTSFVSPKWVPMQGDHTEVLK

dsc          : -----EEEEHHHHHHHH--EHHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--HHHHHHH--HH
jalign      : -----EEEEEE-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--EEEE-----HHHHHHHHHHHHHH
jfreq       : -HHHHHHH--HHHHHHHH-----EEEEEE-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--EEEE-----HHHHHH
jhmm        : -----EEEEEE-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--EEEE-----HHHHHHHHHHHHHH
jnet        : -HHHHHHH--HHHHH-----EEEEEE-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--EEEE-----HHHHHHH
jpssm       : -HHHHHHHHHHHHHHH--EEEHH-----EEEEEE-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--EEEE-----HHHHHH
mul         : -----EEEEEE-----H-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--EEEE-----HHHHHHH--HH
phd         : -----EEEEEE-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--EEEEEE-----HHHHHHHHHHHEE-
pred        : -----HHHHHH-----EEEEEE-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--EEEE-----HHHHHHHHHH

Jpred       : -----EEEEEE-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH--EEEE-----HHHHHHHHHH

PHDHtm      : -----
MCoil       : -----
MCoilDI     : -----
MCoilTRI    : -----
Lupas 21    : -----
Lupas 14    : -----
Lupas 28    : -----

PHDacc      : -----BBBBBB-B-U-BB-BBBBBBBB-B--BBBBB-BBB-----B--BBB-B--B-BB-BB--BBB-BB-BBB-BBB--BB-BB--BBB--
Jnet_25     : ---B---B---BBBB-BB-----B-BBBBBBBB-BBB-B-B-B--BB-BB-BBB-BBB-BBBBBBBBBB-BBB-BB--BBB-
Jnet_5      : -----B-B-BBB--B-----B--BB--B--BB--BBB-BBB--B-----
Jnet_0      : -----B-----B-----B--B-----

PHD Rel     : 99899888877777777877887666677872799851456776666432345668999999855982799615557852234565432102
Pred Rel    : 0070770670588670565755079007898690896666787777687556889999999886996886587898998667867888888
Jnet Rel    : 87898861421331001112453415688874799970468888677873658899999998782798186357765433101221589999
```

Ab initio Prediction of Protein Structure

- Need to find a potential function where

$$E(S, C_{\text{native}}) < E(S, C_{\text{non-native}}).$$

- Need to construct an algorithm to find the global minimum of this function.

Still an unsolved, computationally demanding problem

→ **Homology Modeling**

→ **BLAST / PDB**

(find related proteins whose structures are known)



CASP 10

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Credit: 216,670
RAC: 0

Message

[Message 72941](#) - Posted 30 Apr 2012 19:37:35 UTC

Hello everyone !

CASP 10, a community wide experiment in structure prediction starts tomorrow on May 1st and runs to August 1st. During this time we will be using BOINC heavily for structure prediction. If your work unit starts with the label rb you're running a CASP 10 target! rb is short for Robetta which is our publicly available server for structure prediction.

CASP

CASP is an international experiment to assess the state-of-the-art of the protein structure prediction field. Sequences, whose structures have been solved but which have not yet been published are sent out to participating teams and we have a 3 days to send back predictions. The whole thing is conducted in a double-blind fashion ensuring fair assessment and truly blind prediction.

Robetta

Structure prediction for the community, by the community. Robetta is a server for protein structure prediction that shares Rosetta's structure prediction capabilities to the scientific community (and to the public). The computation for this will be conducted on BOINC meaning that you guys will be crunching



C
A
S
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10



- Celebrating 10 CASP experiments
- Tracking nearly 20 years of progress in protein structure modeling
- The latest assessment of the state of the art

Retrospective and prospective views of the field from



Joel Sussman

Gaeta, Italy

December 9 -12, 2012

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Welcome to the Protein Structure Prediction Center!

Our goal is to help advance the methods of identifying protein structure from sequence. The Center has been organized to provide the means of objective testing of these methods via the process of blind prediction. The Critical Assessment of protein Structure Prediction (CASP) experiments aim at establishing the current state of the art in protein structure prediction, identifying what progress has been made, and highlighting where future effort may be most productively focused.

There have been nine previous CASP experiments. The tenth experiment is planned to start in April 2012. Description of these experiments and the full data (targets, predictions, interactive tables with numerical evaluation results, dynamic graphs and prediction visualization tools) can be accessed following the links:

[CASP1 \(1994\)](#) | [CASP2 \(1996\)](#) | [CASP3 \(1998\)](#) | [CASP4 \(2000\)](#) | [CASP5 \(2002\)](#) | [CASP6 \(2004\)](#) | [CASP7 \(2006\)](#) | [CASP8 \(2008\)](#) | [CASP9 \(2010\)](#) | [CASP10 \(2012\)](#)

Raw data for the experiments held so far are archived and stored at our [data archive](#).

Starting November 2011, we are opening a new rolling CASP experiment for all-year-round testing of ab initio modeling methods:

[CASP ROLL](#)

Details of the experiments have been published in a scientific journal *Proteins: Structure, Function and Bioinformatics*. [CASP proceedings](#) include papers describing the structure and conduct of the experiments, the numerical evaluation measures, reports from the assessment teams highlighting state of the art in different prediction categories, methods from some of the most successful prediction teams, and progress in various aspects of the modeling.

Message Board

Sep.6 - early bird registration deadline; CASP fellowships letters

[Dear CASP participants, All recipients of CASP student fellowships have been identified and notified. Please proceed with the registrations according to the instructions provided in the award let ...](#)

Abstract collection; meeting fellowships; early bird registration

[1. Just three days ago we received the last CASP10 prediction, and today we start collecting methods abstracts. The Abstract Submission web page is available through a link from the CASP10 main page. ...](#)

End of CASP10 regular