### Sedimentation velocity



Sedimentation velocity

- High rotor velocity
- Long solution column
- Hydrodynamic information



### Sedimentation velocity analysis

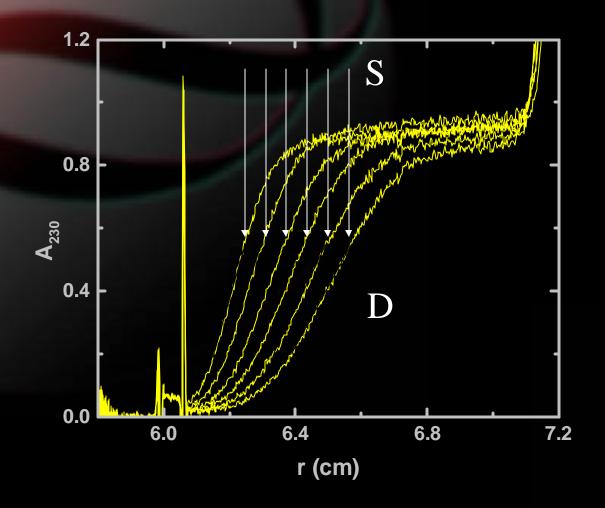
Simple theory

Effects of hydrodynamic and thermodynamic nonideality

Different methods of analysis

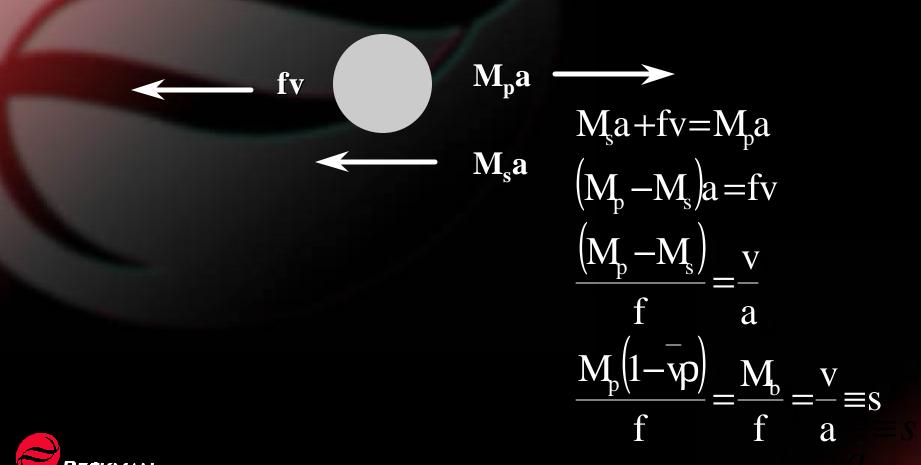


### Sedimentation velocity





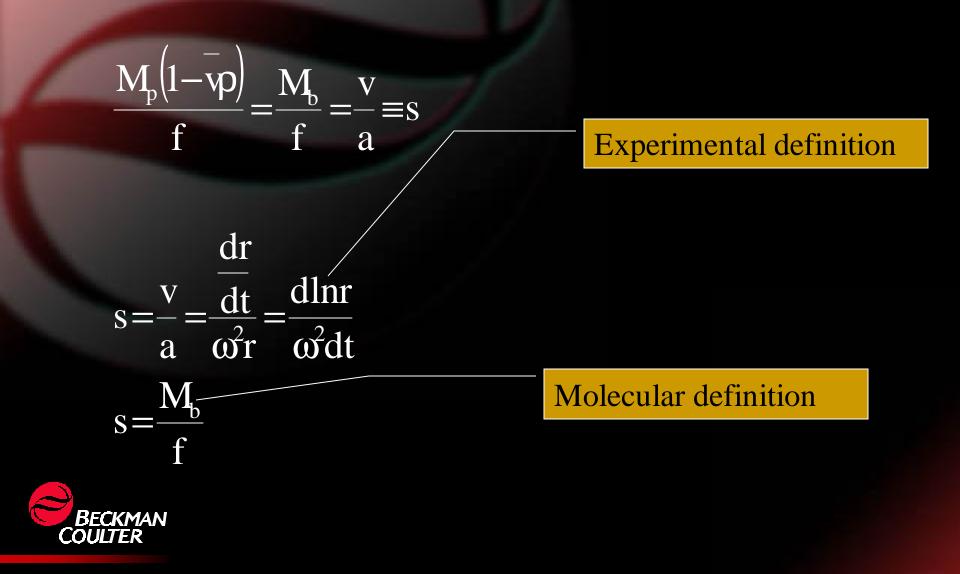
## Simple theory: Analysis of forces



V



#### Velocity theory-continued



Number of species

Aggregate test

Sedimentation coefficient

Shape/hydration



## Preliminaries

What do you want to know?

Sample handling

Sample type

Optical system



#### Sedimentation Velocity

**Select Operating Conditions** 

Select rotor speed Select temperature Select optical system

General requirements

450 uL sample/cell C depends on optical system

Select Method of Analysis

g(s) good for distributions Transport equation (e.g.Svedberg) good for low M Sw vs. [c] association constants Van Holde-Weischet good for pauci-disperse systems



## Notes on Sedimentation velocity

Try to run 3 or more concentrations

From highest -> optical system limit

 For total unknown start run at low rotor speed (3000 rpm)

 Run at multiple rotor speeds or use gravitational sweep



#### Preliminaries

What do you want to know?

Sample handling

Sample type

Optical system



# Sample type

<u>Protein</u>	Polysaccharide	Nucleic Acid
Choice of optics	Interference optics	Absorbance optics
1 A <sub>230 or 280</sub>	C > 1 mg/ml	1 A <sub>260</sub>
1 mg/ml	Nonideality	Nonideality



#### Preliminaries

What do you want to know?
Sample handling
Sample type
Optical system



## Choosing optical system

#### Use absorbance if:

- Need selectivity
- Added sensitivity
- Cannot dialyze sample

#### Use both:

- Determine extinction coefficient
- Test for sample purity
- Extend concentration range

#### Use interference if:

- Buffer absorbs
- Sample does not absorb
- Precision required
- g(s)
- Extinction coefficient varies
- Short columns



Number of species

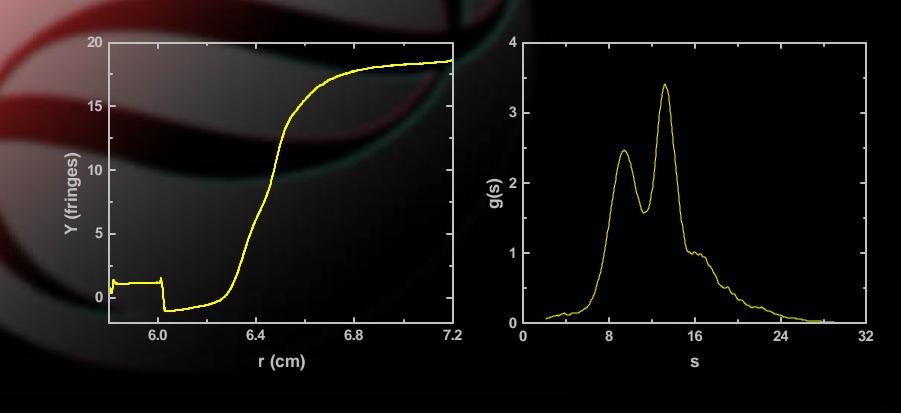
• Aggregate test

Sedimentation coefficient

Shape/hydration



## Number of species



Raw data





Number of species

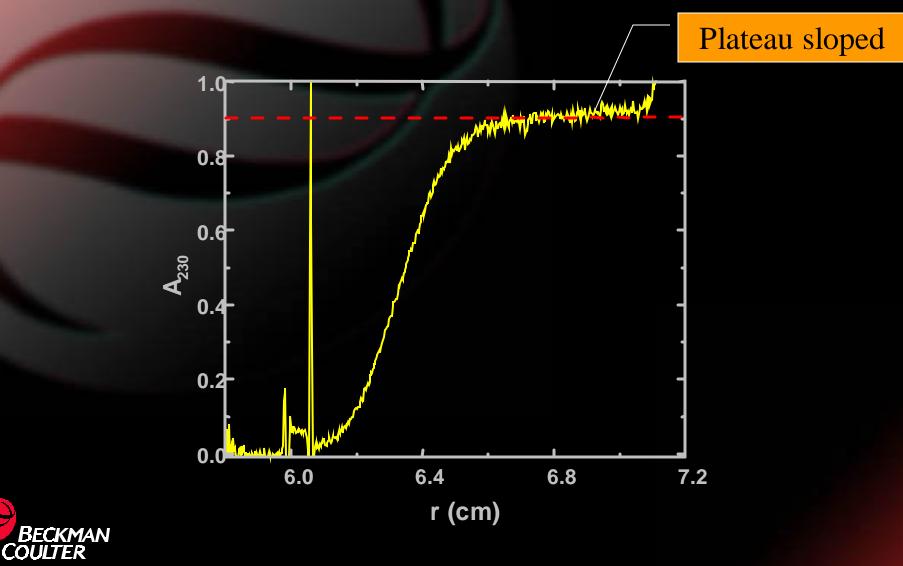
• Aggregate test

Sedimentation coefficient

Shape/hydration







Number of species

Aggregate test

Sedimentation coefficient

Shape/hydration

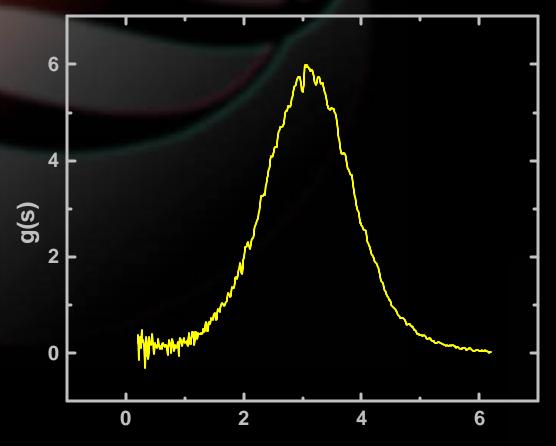


#### Determining s

Whole boundary methods
Transport equation
Fitting to simulation
Time derivative
van Holde Weischet



#### Determining s Time derivative method





Number of species

• Aggregate test

Sedimentation coefficient

Shape/hydration



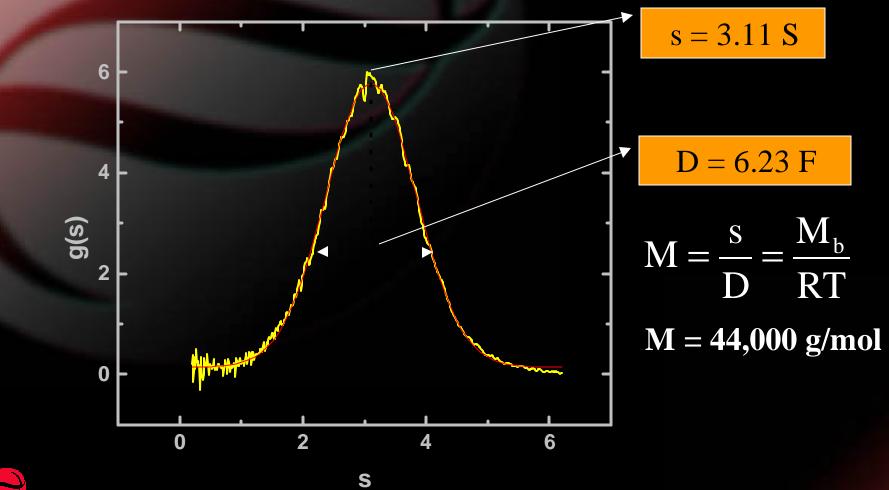
Number of species

Aggregate test

Sedimentation coefficient

Shape/hydration







#### General considerations

Correcting for buoyancy
 Determining density
 Partial specific volume
 Correcting for viscosity



#### Useful references

#### **Books:**

Analytical Ultracentrifugation in Biochemistry and Polymer Science. (1992) S.E. Harding, A.J. Rowe, and J.C. Horton, eds. Royal Society of Chemistry, Cambridge.

Modern Analytical Ultracentrifugation. (1995) T.M. Schuster and T.M. Laue, eds. Birkhauser, Boston. *Two fairly recent books devoted entirely to this field* 

K.E. van Holde, <u>Physical Biochemistry</u>. (1985) Prentice Hall, Englewood Cliffs, New Jersey. Good introductory text for general theory of sedimentation, frictional coefficients, diffusion, and other hydrodynamic analysis

Freifelder, D. (1982). Physical Biochemistry: Applications to biochemistry and molecular biology. W.H. Freeman, New York.

Regarded as a good introductory text that is strong on centrifugation methods

van Holde, K.E., W.C. Johnson, Jr., and P.S. Ho. .(1998). Principles of physical biochemistry. Prentice-Hall, Upper Saddle River.

Cantor, C.R. and Schimmel, P.R. (1980). Biophysical chemistry. Part II: Techniques for the study of biological structure and function. W.H. Freeman, San Francisco. *These two are more advanced texts with good coverage of centrifugation methods* 

#### **Special Journal Issue:**

*Chemtracts Biochemistry and Molecular Biology*, vol. 11 no. 13 (pp. 933-1004), December 1998 (Jeffrey C. Hansen, Guest Editor) *Several review articles and condensation commentaries on current research* 



#### Useful references

#### **Review Articles:**

Stafford, W.F. III. (1997). Sedimentation velocity spins a new weave for an old fabric. *Curr. Opin. Biotechnol.* 8, 14-24.

Laue, T.M. (1995). Sedimentation equilibrium as thermodynamic tool. Methods Enzymol. 259, 427-452.

Laue, T.M. Stafford, W.F., III (1999). Modern Applications of Analytical Ultracentrifugation. *Annu. Rev. Biophys. Biomol. Struct.* 28, 75-100.

#### **Articles:**

Laue, T.M., Shah, B.D., Ridgeway, T.M., and Pelletier, S.L. (1992). Computer-aided interpretation of analytical sedimentation data for proteins. In: Analytical ultracentrifugation in biochemistry and polymer science. S.E. Harding, A.J. Rowe, and J.C. Horton, eds. Royal Society of Chemistry, Cambridge, pp. 90-125. *Procedures for calculating partial specific volume, density, sedimentation coefficient (corrected for water @ 20°C and extrapolated to zero concentration) hydration, frictional rations, ellipsoidal shapes, etc; basis for SEDNTERF software (but note that the formulae and tables contain a number of typographical errors that were corrected in SEDNTERP – see the SEDNTERP Help file for corrected formulas)* 

Stafford, W.F., III. (1992). Boundary analysis in sedimentation transport experiments: A procedure for obtaining sedimentation coefficient distributions using the time derivative of the concentration profile. *Anal. Biochem.* 203, 295-301. *Initial publication describing the dc/dt method* 

Johnson, M.L. and Frasier, S.G. (1985). Nonlinear least-squares analysis. *Methods Enzymol*. 117:301-342. *Good overview of the fitting of experimental data* 

